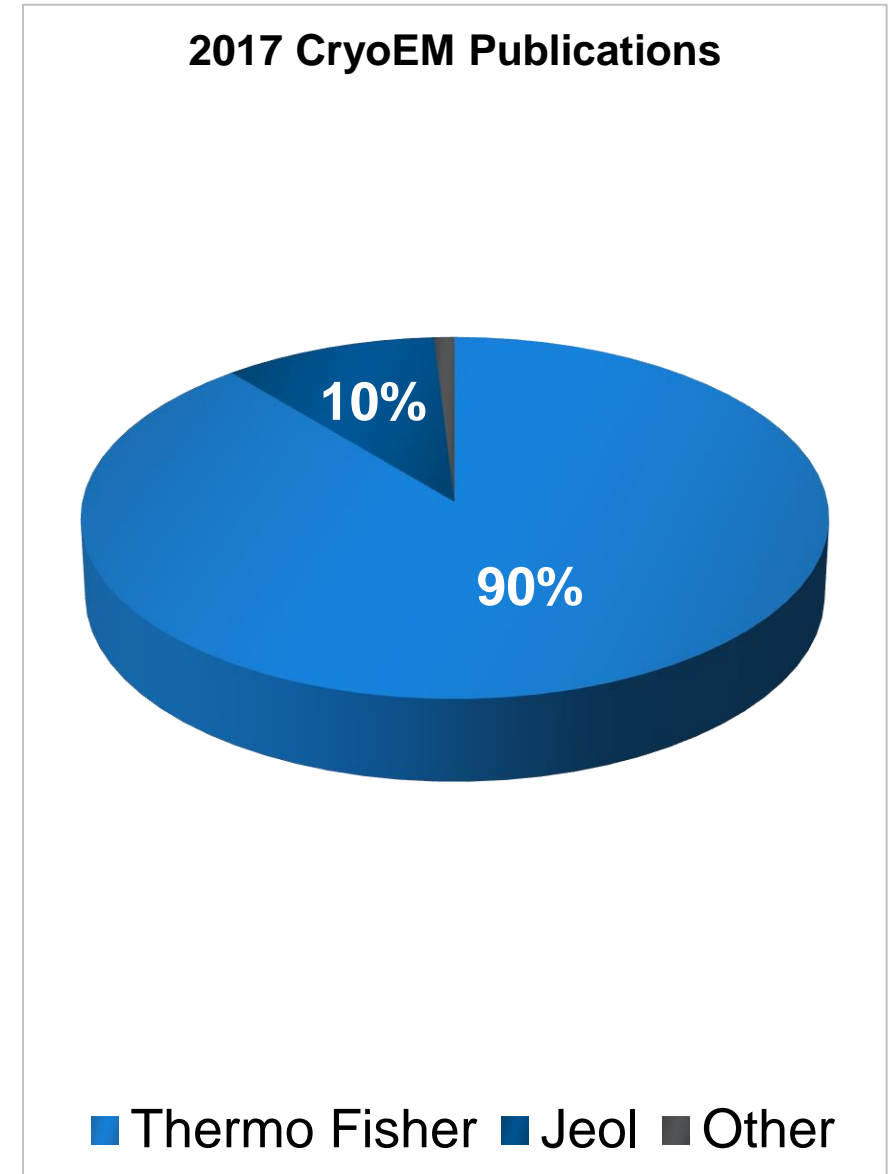
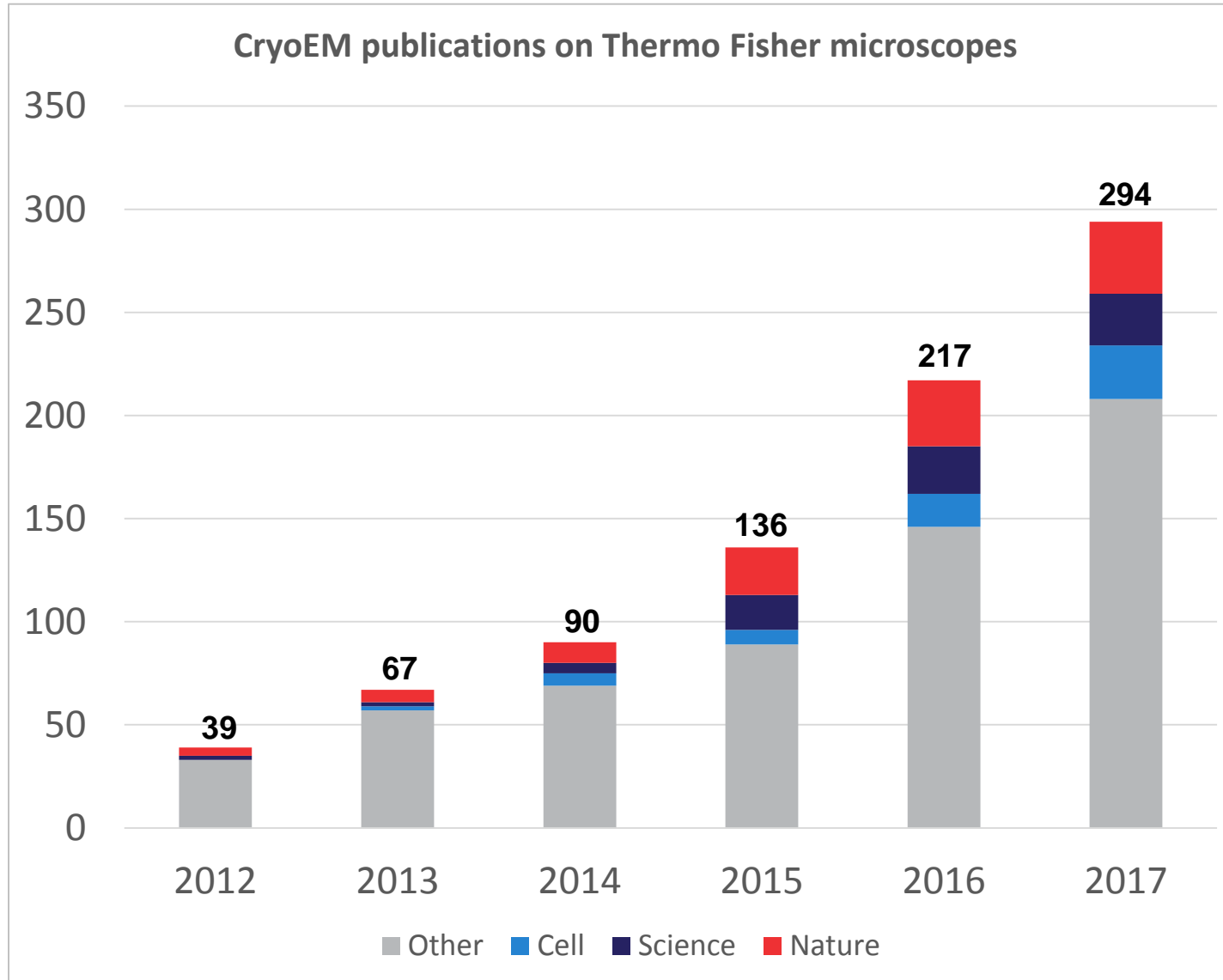




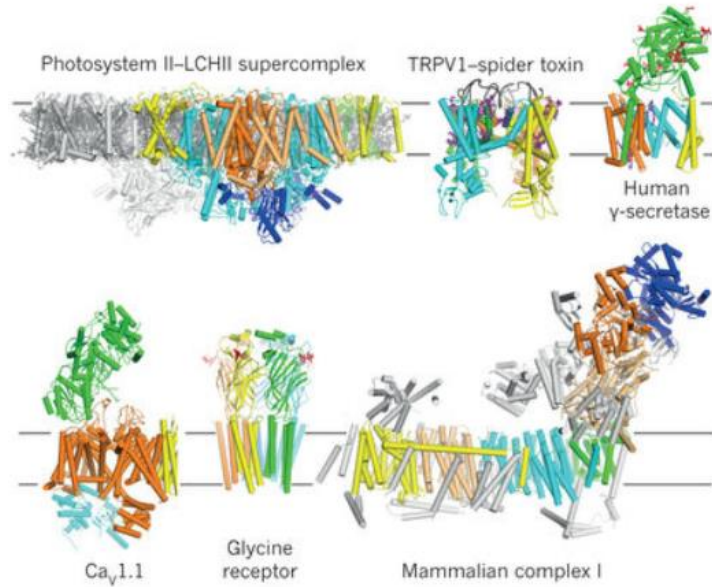
Single Particle Analysis (SPA) Workflow
Understanding molecular structures – faster and more reliably with CryoEM

Medinprot, Budapest
21-04-2018
Max Maletta

Growth in structural biology CryoEM over the past 5 years

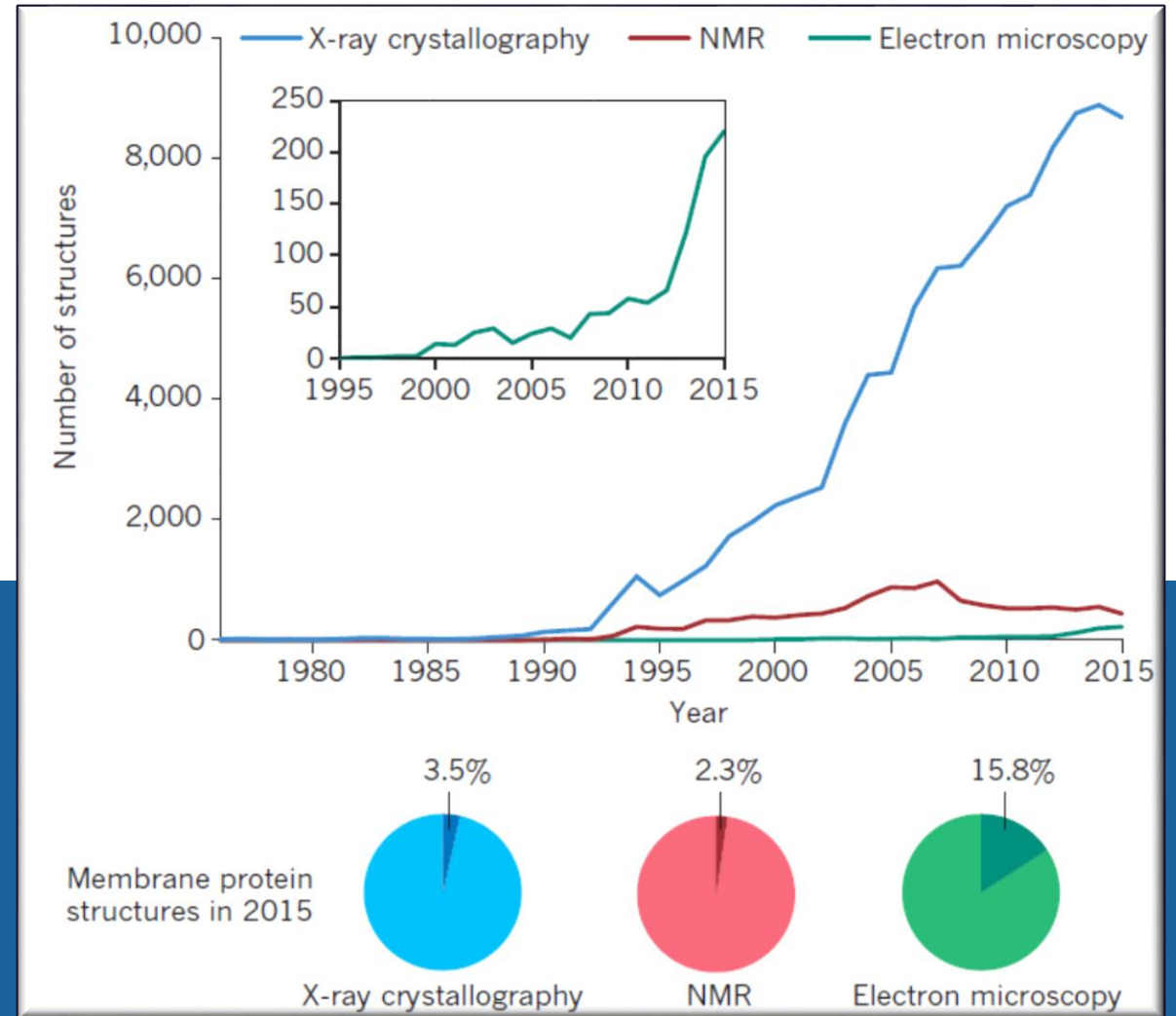


Growth in structural biology over the past 40 years



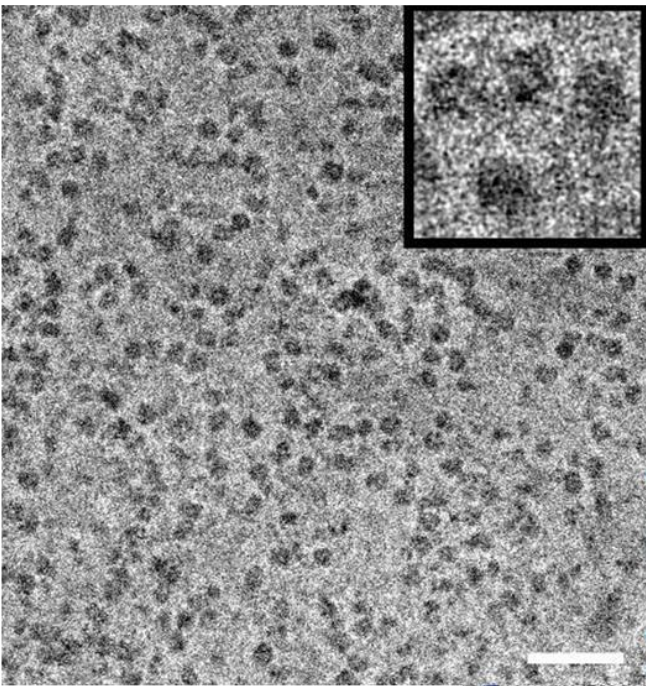
Examples of membrane protein structures that were determined through cryo-EM

“Developments in the electron microscopy are providing unprecedented opportunities for the structural characterization of biological macromolecules. This is resulting in a wave of information about processes in the cell that were impossible to characterize with existing techniques in structural biology.”

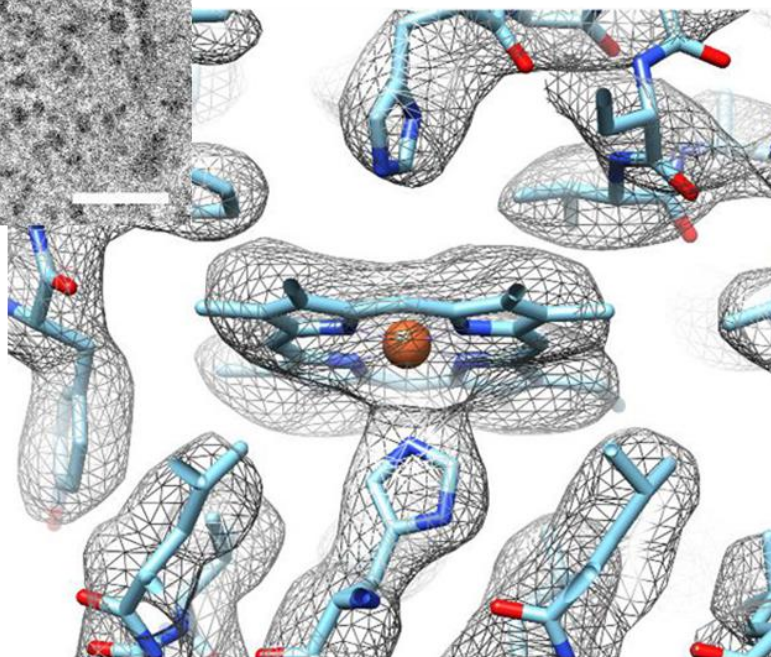


R. Fernandez-Leiro & S. H. W. Scheres Nature 2016

Resolution revolution from “blob-ology” to atomic resolution



Titan Krios with Volta Phase Plate was tested for the most challenging small particles – hemoglobin (**MW 64 kDa** and C2 symmetry)



Cryo-EM structure of haemoglobin at 3.2 Å determined with the Volta phase plate
bioRxiv preprint 2016
Submitted to Nature 2016
M. Khoshouei, M. Radjainia, W. Baumeister & R. Danev

Titan Krios TEM 300kV

- Proven performance $< 2\text{\AA}$
 $< 100\text{kDa}$



Glutamate dehydrogenase

- 334 kDa
- **1.8Å resolution**

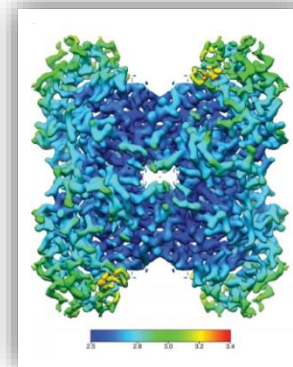
Breaking Cryo-EM Resolution Barriers to Facilitate Drug Discovery
Cell, 2016

A.Merk, A. Bartesaghi, S. Banerjee, L. A. Earl, J. Milne, S. Subramaniam



Talos Arctica TEM 200kV

Proven performance $< 3\text{\AA}$



Rabbit muscle aldolase

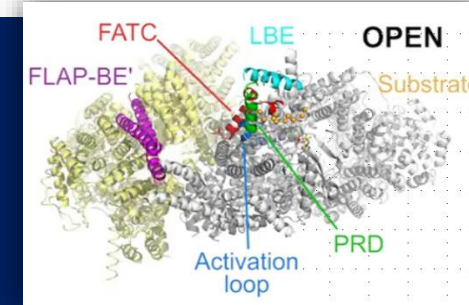
- 150 kDa
- **2.6Å resolution**



Merk, A. et al. (2016), *Cell*, 165: 1698
Herzik, M.A. Jr et al (2017), *preprint on biorxiv*, <http://dx.doi.org/10.1101/141994>

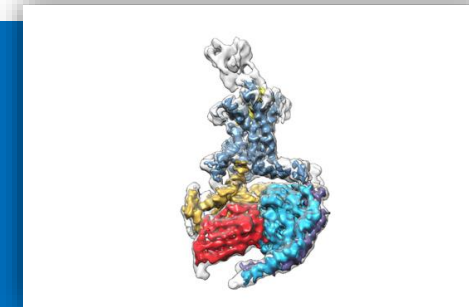
CryoEM SPA is complementary to traditional structural biology techniques

Study of **dynamic** biological processes



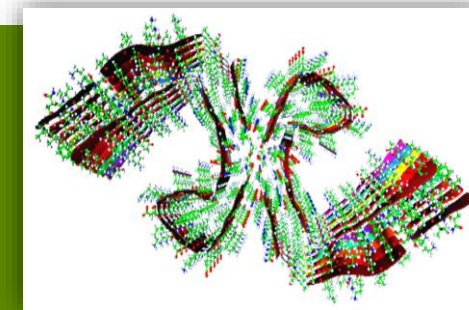
Human ATM
Multiple conformations

Structures that are difficult to solve with other techniques: **membrane** proteins and ion channels

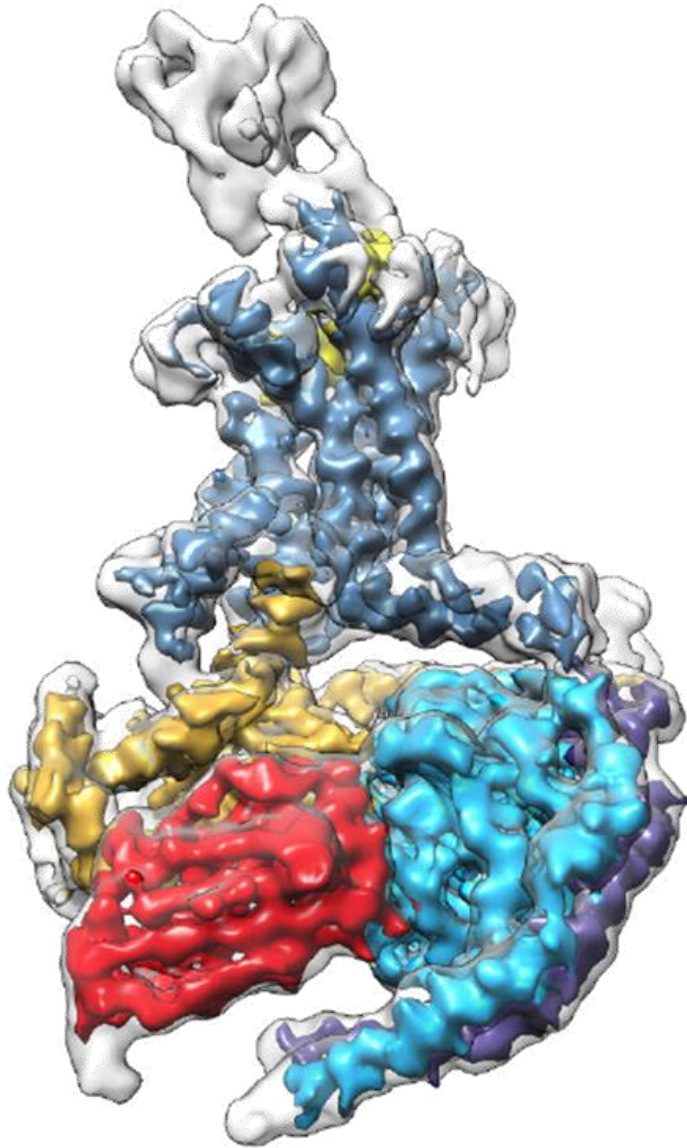


GPCRs
Large membrane protein

Atomic resolution structures of protein **complexes**, aggregates and large virus assemblies



TAU protein
Aggregation of a protein complex



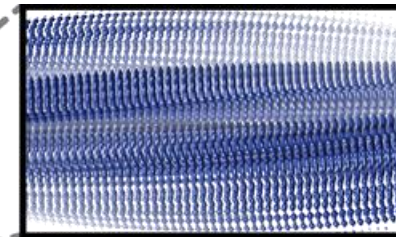
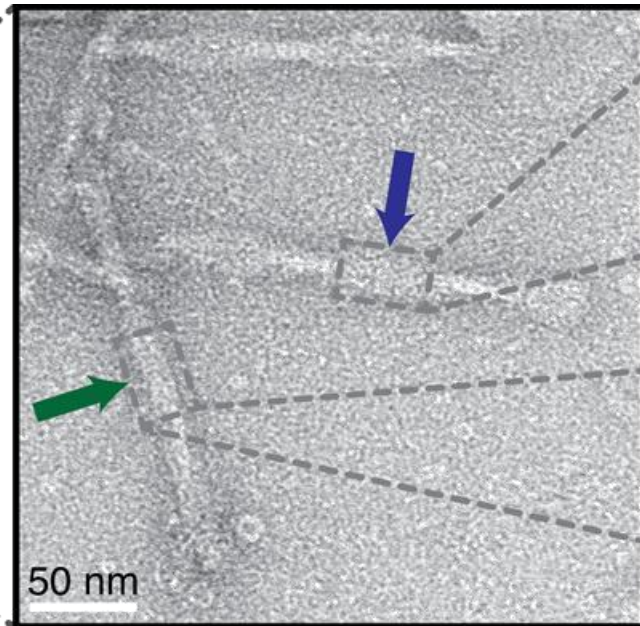
- G protein-coupled receptors (GPCRs) are major targets for treatment of chronic diseases
- Research objective
Understand mechanism of membrane trafficking – “functional states”
- Unique
CryoEM allows to visualize the structure of an *activated* GPCR complex (bound to proteins)
- **This information supports the design of better drugs to treat diabetes, obesity, osteoporosis and migraine**

CryoEM of Tau protein filaments provides insight in aggregation mechanism

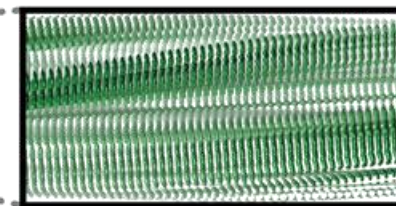
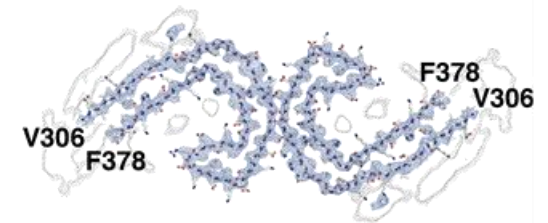
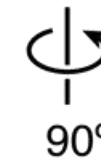
complex

- Alzheimer's disease caused by Tau protein filaments aggregating in the brain (protein stacks → plaque)
- Research objective
Understand mechanism of protein aggregation
- Unique
CryoEM revealed structure of 2 distinct types of filament – small tweaks in how monomers associate with one another has drastic effects on the fibril shape
- **This understanding of the aggregation mechanism contributes to future drug design**

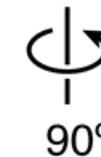
Alzheimer's disease (AD) brain from deceased patient



Cryo-EM structure of Tau Paired Helical Filament



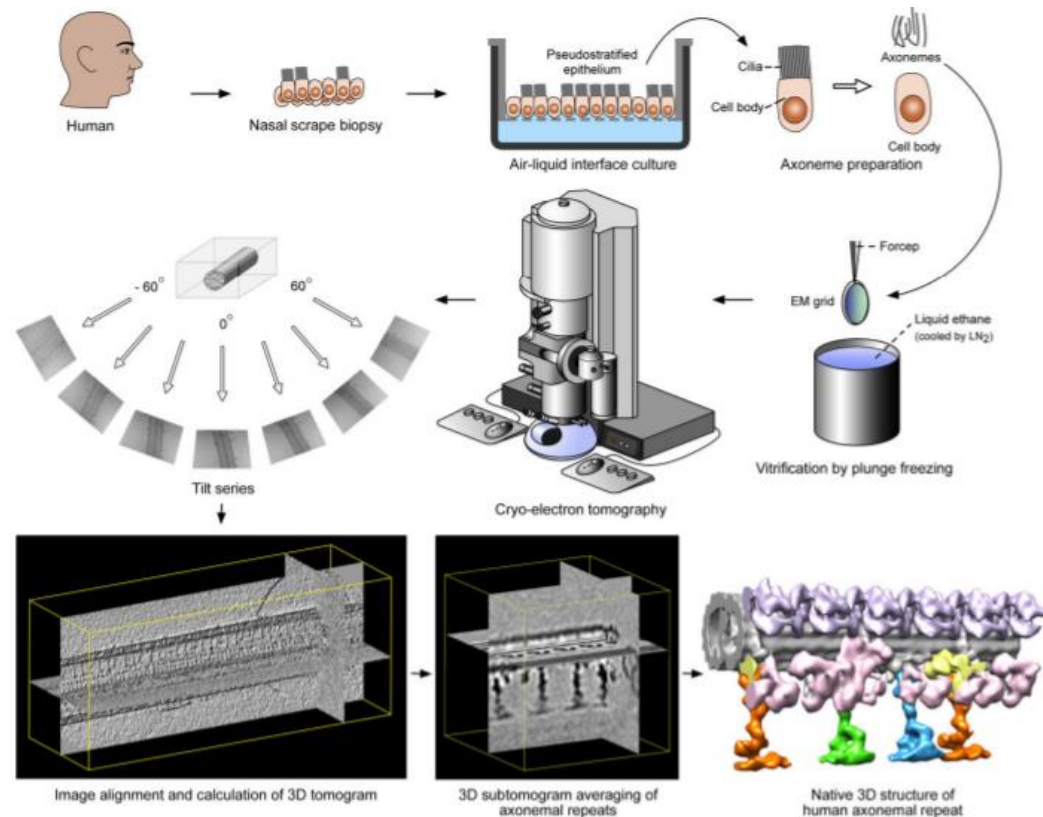
Cryo-EM structure of Tau Straight Filament



Fitzpatrick et.al. (2017) *Nature* **547**, 185-190 | doi:10.1038/nature23002

Cryo-electron tomography reveals ciliary defects underlying human *RSPH1* primary ciliary dyskinesia

Jianfeng Lin¹, Weining Yin², Maria C. Smith¹, Kangkang Song¹, Margaret W. Leigh⁵, Maimoona A. Zariwala³, Michael R. Knowles⁴, Lawrence E. Ostrowski^{2,*}, and Daniela Nicastro^{1,*}



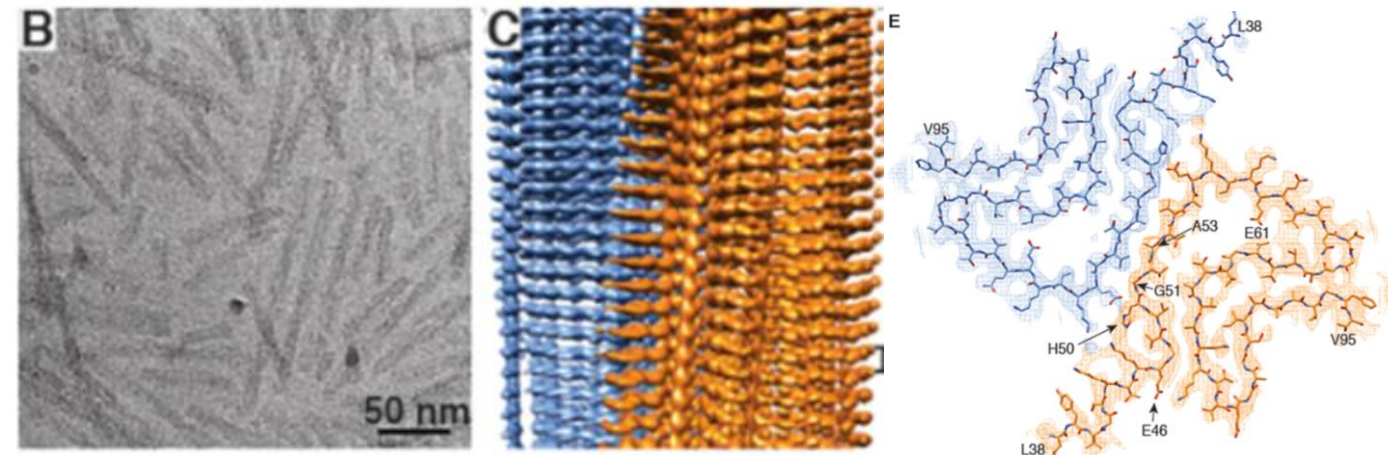
STRUCTURAL BIOLOGY

Fibril structure of amyloid- β (1-42) by cryo-electron microscopy

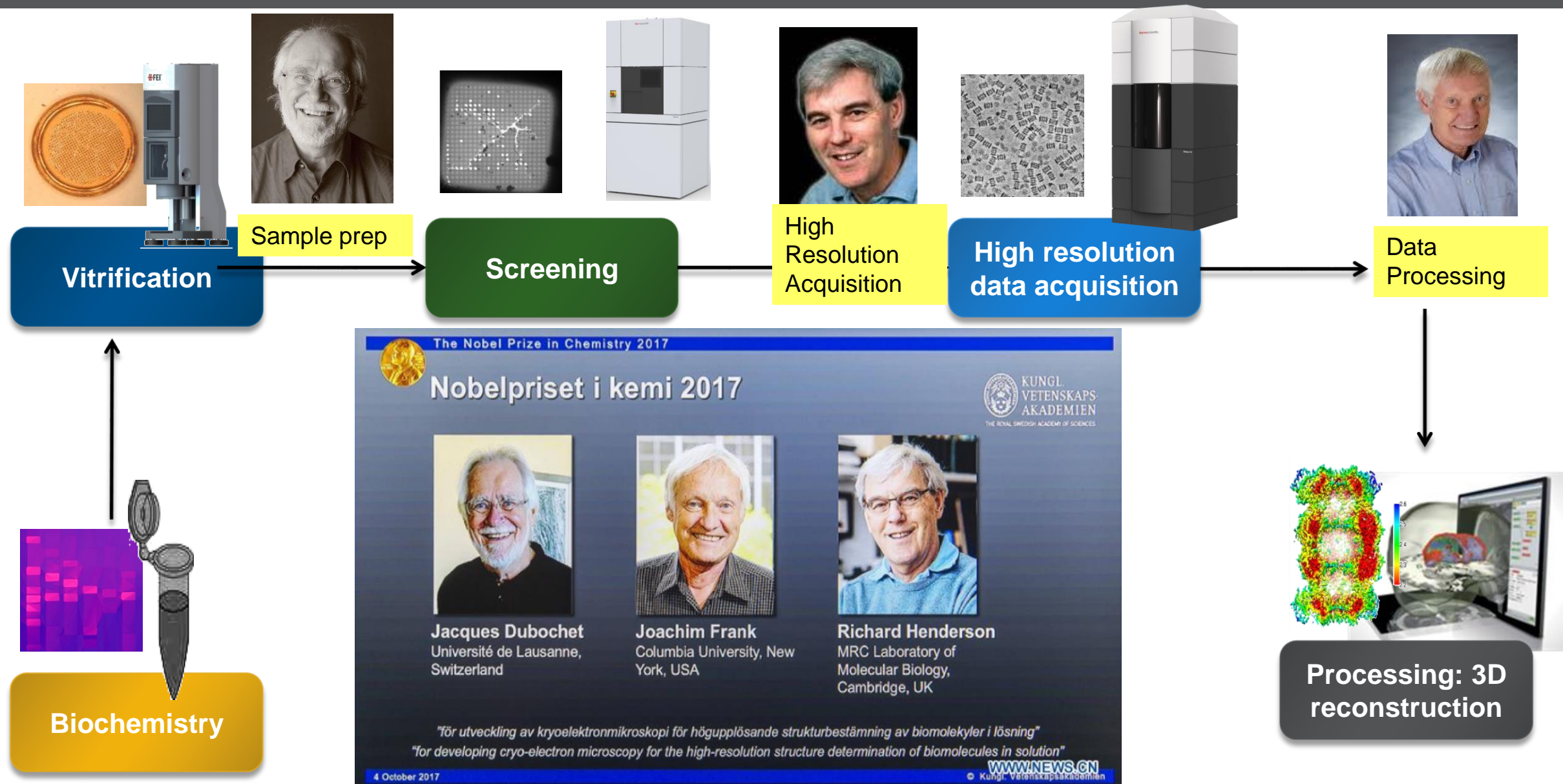
Lothar Gremer,^{1,2} Daniel Schölzel,^{1,2} Carla Schenk,¹ Elke Reinartz,² Jörg Labahn,^{1,2,3} Raimond B. G. Ravelli,⁴ Markus Tusche,¹ Carmen Lopez-Iglesias,⁴ Wolfgang Hoyer,^{1,2} Henrike Heise,^{1,2} Dieter Willbold,^{1,2*} Gunnar F. Schröder^{1,5*}

Cryo-EM structure of alpha-synuclein fibrils

Ricardo Guerrero-Ferreira¹, Nicholas M.I. Taylor^{1†}, Daniel Mona², Philippe Ringler¹, Matthias E. Lauer³, Roland Riek⁴, Markus Britschgi², and Henning Stahlberg^{1,*}



2017 Breaking News: Nobel Prize for CryoEM SPA Workflow

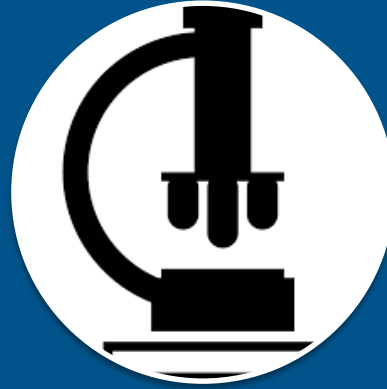


Single Particle Analysis (SPA) Workflow Challenges



Biochemistry and vitrification complexity

- Efficient and affordable screening & sample optimization



Reproducibility of best performance

- Ease-of-use through extensive automation



Access to expertise and productivity

- Customized systematic approach to integrated service and applications support



New in the Single Particle Analysis (SPA) Portfolio



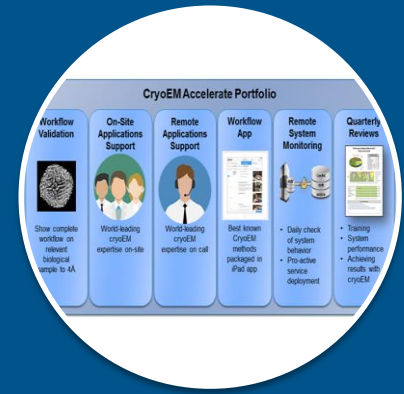
Glacios Cryo-TEM 200 kV FEG + AutoLoader

- Efficient and affordable screening solution



Krios G3i Cryo-TEM 300 kV FEG + AutoLoader

- Ease-of-use through extensive automation



Accelerate portfolio for customer success

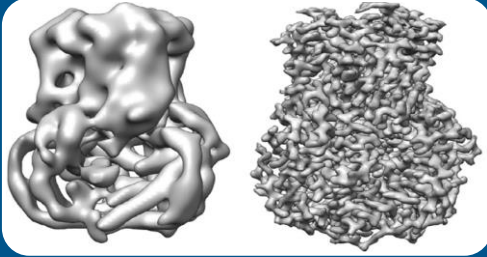
- Individualized systematic approach to integrated service and applications support

Access the next level state-of-the-art cryo electron microscopy, with improved ease-of-use and performance

- Optimal tool performance always guaranteed
- Automation and guidance simplifies experiment set up
- Highest resolution data at maximum throughput



Krios G3i Cryo-TEM – Workflow Performance Improvements



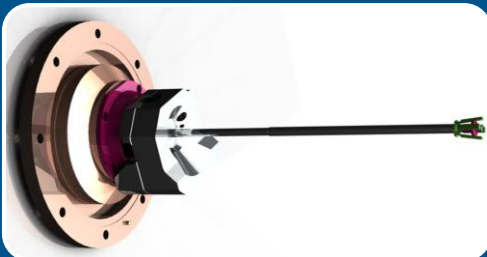
Data quality → 3D model resolution

- Guaranteed low anisotropic magnification distortion: < 1%
- Magnification range: 20kX → 150 kX



Throughput

- Improved drift after transfer
- Atlas speed improvement: immediately after transfer and data acquisition within minutes
- Eucentricity improvement for tomography: 1x1x3 μm^3



Sample life time

- Improved cryo box with anti contaminator
- 3 days guaranteed (column)
- 5 days in Autoloader

Grant, T. et al. (2015), *J. Structural Biology*, 192(2): 204

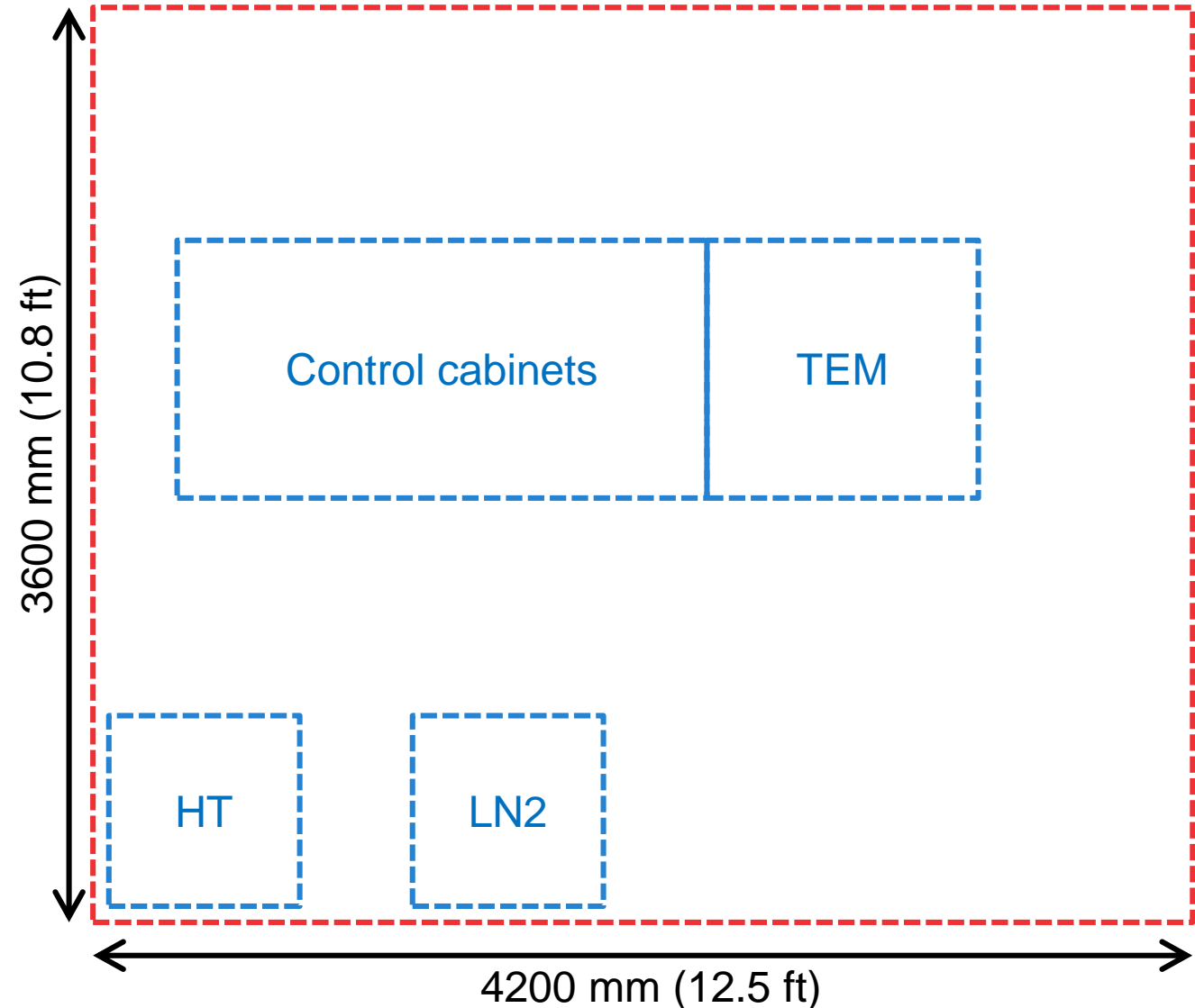
- *Throughput acceleration by efficient cryo-screening of particle distribution and ice quality*
- *An affordable and complete solution for routine SPA data acquisition*

- Optimal tool performance always guaranteed
- Automation and guidance simplifies experiment set up
- Cryo, Autoloader, 200 kV electron optics on a brand new hardware platform
- Seamless connectivity to Krios TEM and small footprint for reduced installation requirements
- Smaller footprint



Small Footprint and Easy Access for Reduced Glacios Installation Requirements

- New, more modular hardware platform
 - Room floor space: 3800 mm x 3300 mm
 - Room height requirement: 2800 mm (9.2 ft)
-
- Access route width: 900 mm (3 ft)
 - Access route height: 2300 mm (7.6 ft)



Krios G3i and Glacios Cryo-TEMs – Reproducible and Optimal Performance

Data acquisition quality and efficiency

Depend on status of alignments – must always be optimal when starting data acquisition



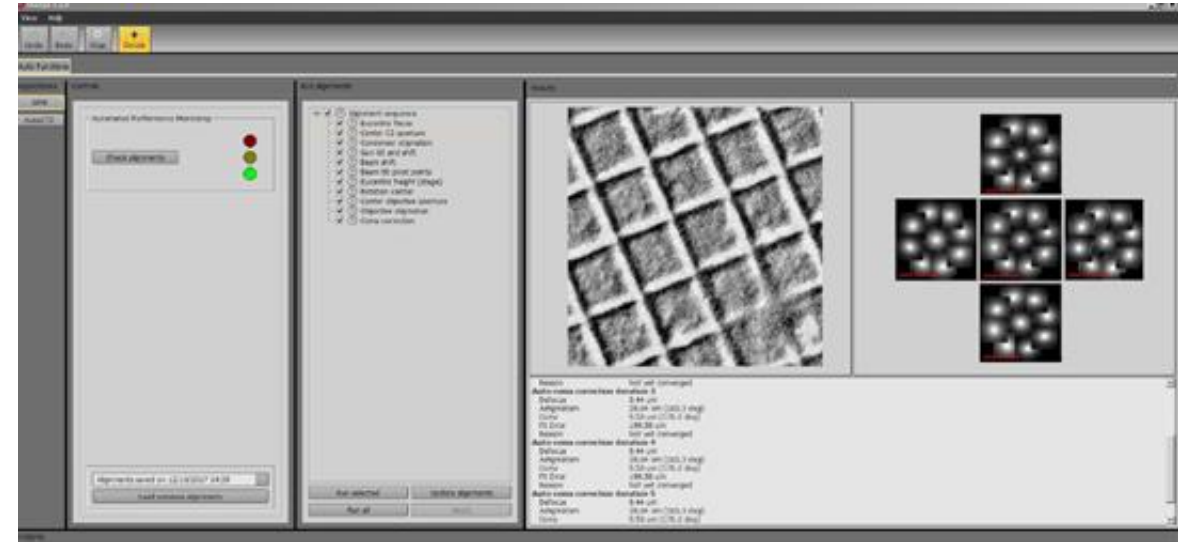
Self-assessment of the actual microscope status
Software for optimization of all alignments

Performance measurement and alignment automation



Reduce alignment complexity for users

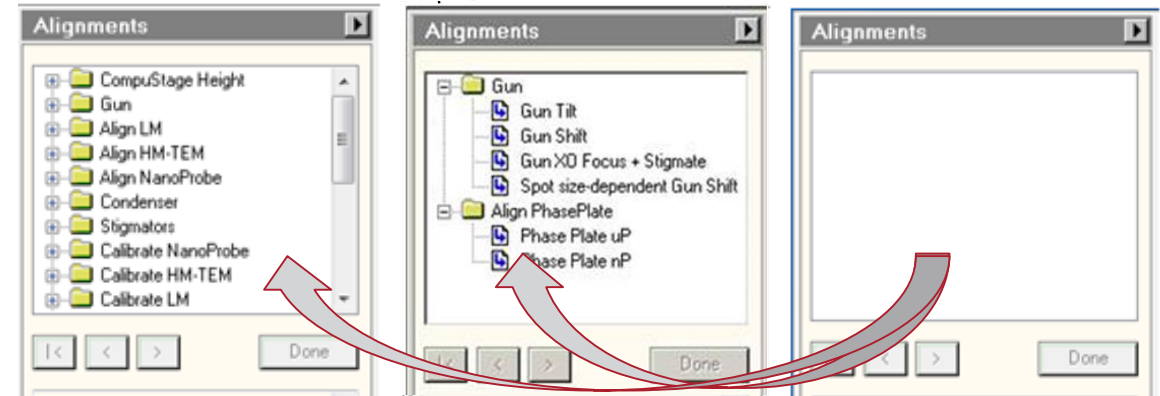
Alignment automation and control access to unnecessary alignments



Factory / Service

Supervisor

User



Krios G3i and Glacios Cryo-TEMs – Simplified Experiment Setup

Full SPA workflow control

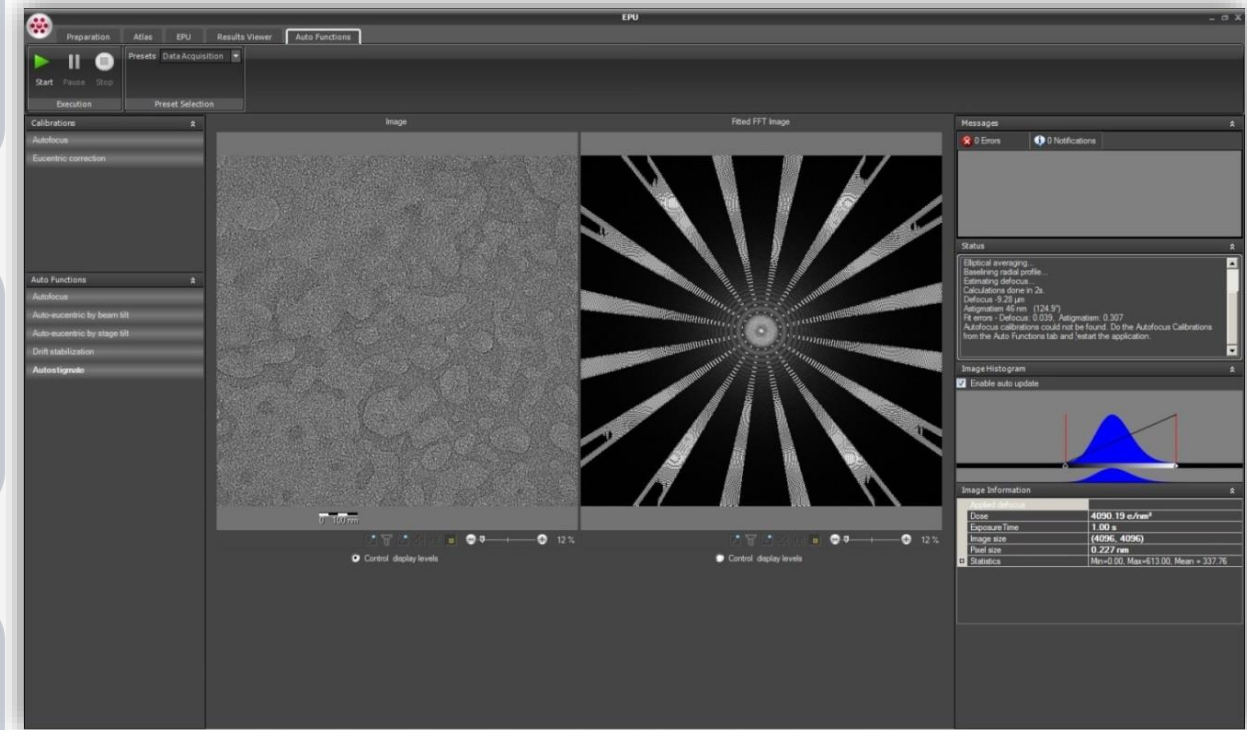
- Microscope setup
- Sample screening
- Data acquisition
- All controlled from EPU UI

One UI for “everything”

- Logical workflow guidance
- Automation in each step

Automated “daily” alignments

- Auto-eucentricity
- Auto-focus
- Auto-coma
- Auto-stigmatize



Automated CTF fitting

Krios G3i and Glacios Cryo-TEMs – Efficient Screening and Data Acquisition

Batch acquisition on all 12 grids

Autoloader control via EPU → automated grid atlas of all grids in a Cassette



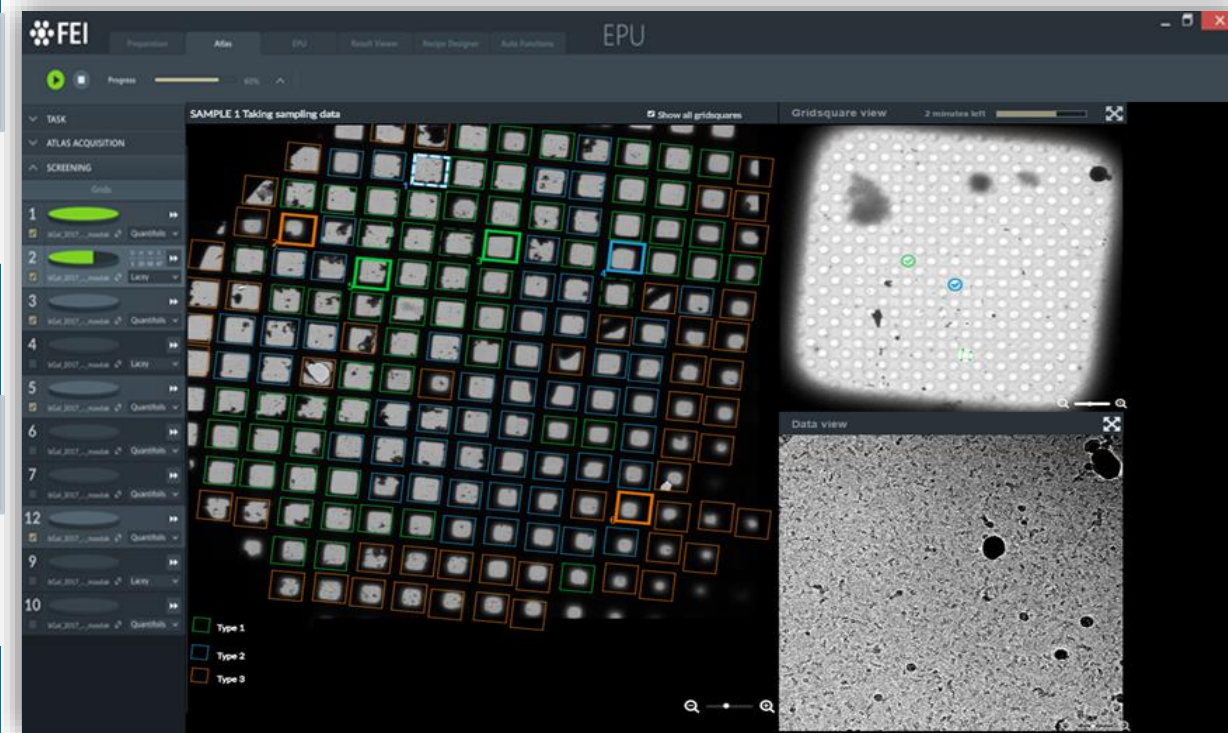
Automated classification of ice

Grid screening on selected grids → automatically analyze ice presence and thickness and classify in groups



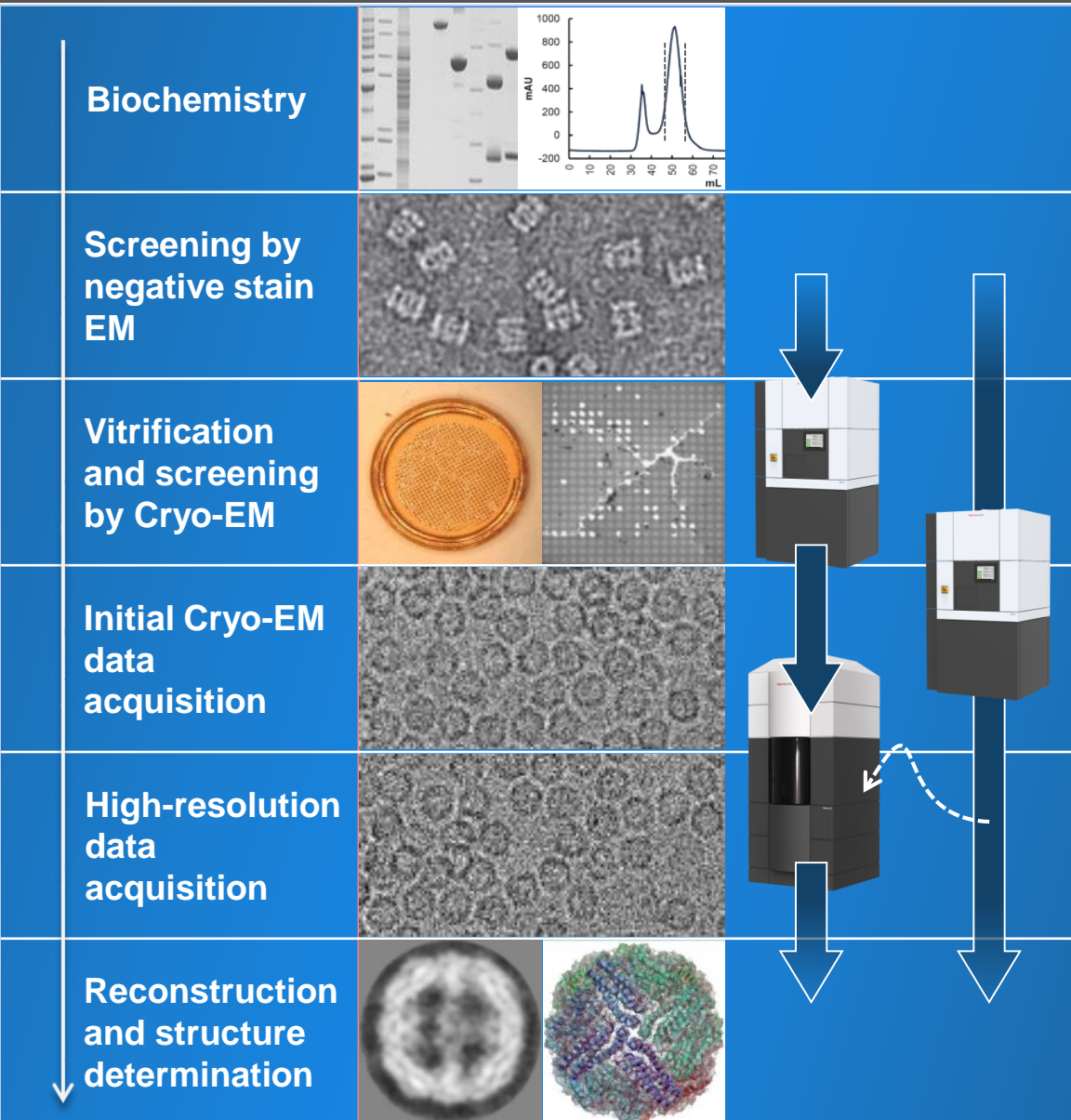
Simplified acquisition of high magnification images for evaluation of ice quality and particle distribution/orientation

Single-click acquisition including autofocus outside the target field of view



Guided selection of grid squares

Connectivity Between Glacios and Krios/Arctica Systems for Improved Workflow Setup

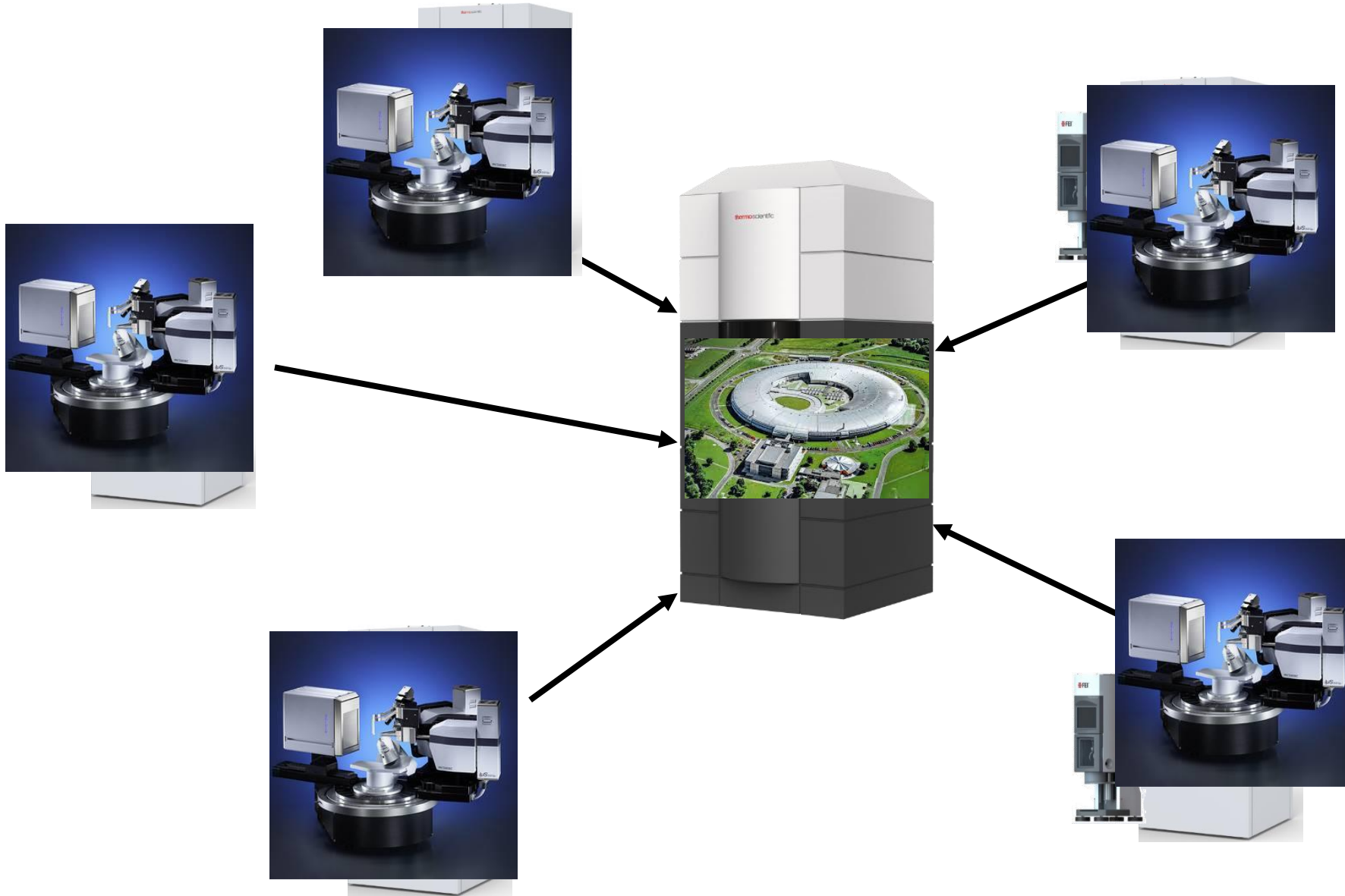


- Contamination- and risk-free transfer
 - Using Autoloader Capsule
 - No manipulation of small parts (grids)
 - No exposure to environment
- Full compatibility:
 - Capsules – Autoloaders – microscopes



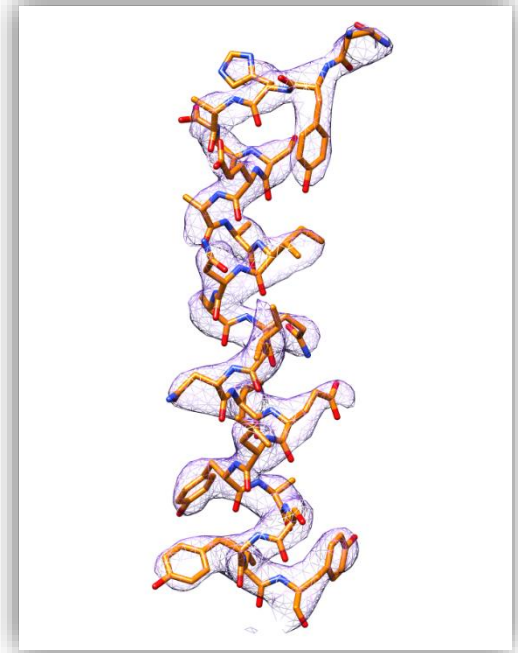
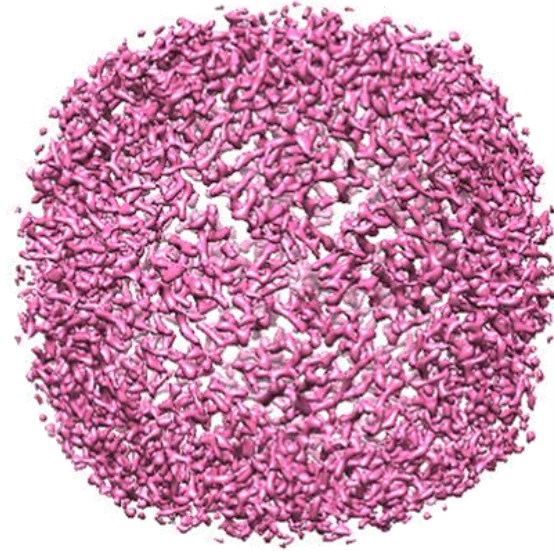
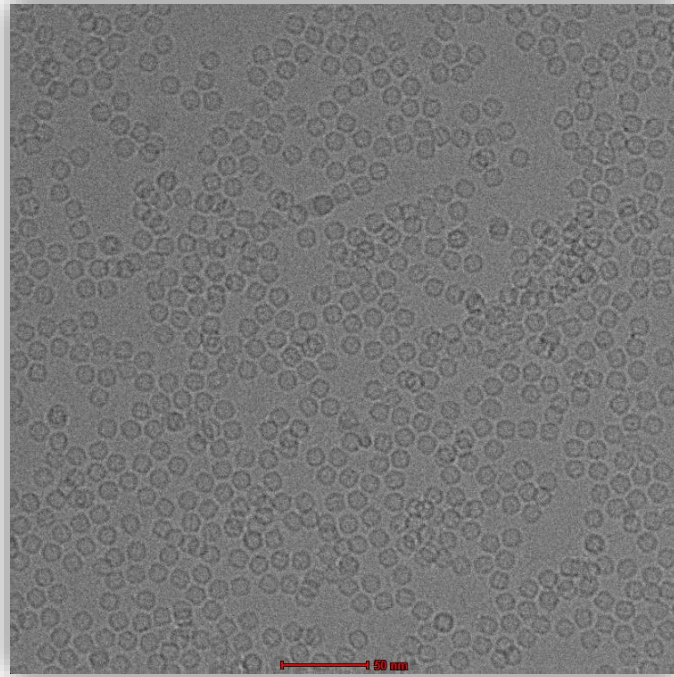
- Prescreening samples in Glacios system → high-resolution data acquisition in Krios system
- Glacios-based laboratories → pass exceptional samples to a Krios system for additional analysis

Cryo-EM adopt the “Synchrotron” model



- Glacios for (cryo)sample quality evaluation and optimization (also room temp negative screening possible)
- Glacios for 2D-classes and initial model 3D model (required for users to get access to centralized facilities' Krios)
- Glacios for high-resolution 3D models (if no ultra-high resolution is required)

Performance: Apoferritin reconstruction at 3.3 Å resolution (Glacios/Falcon 3EC)



Timeline (days)

0

Automated
data collection

~1

Data processing

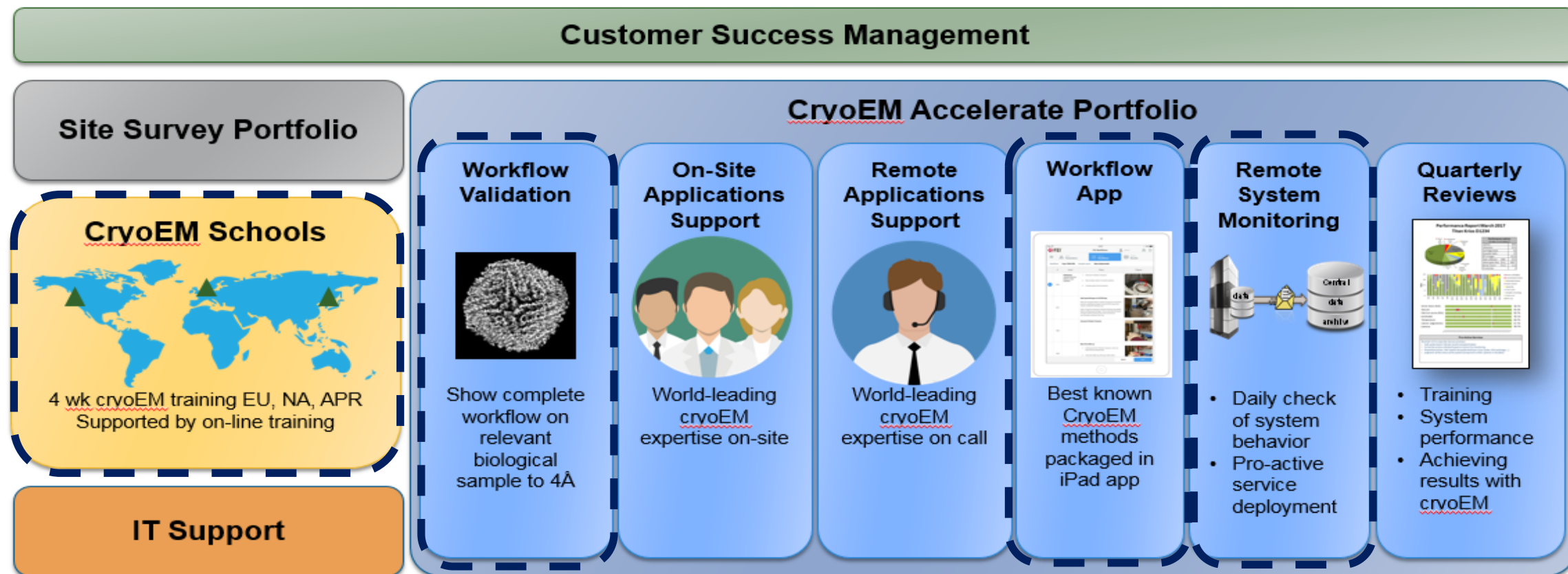
~2

Apoferritin Reconstruction

@ 3.3 Å

Glacios / Falcon 3EC

Personalized Support via Dedicated and Expanding Teams



On-site

- Assistance and training for all workflow components
- Support with the Workflow Assistant App

Remote

- General workflow assistance
- Support with the Workflow Assistant App
- Guided user assistance via RAPID system
- Offline support via email

CryoEM Single Particle Analysis: Summary

- Proven technology, best performance

- >90% of high resolution structures is based on Thermo Fisher microscope data
- Nearly 1020 structures deposited based on FEI microscope data (EMDB 2017)
- Nearly 294 high impact papers based on Titan Krios data (2017)

- Workflow connectivity

- 200+ installed base (Titan Krios and Talos Arctica)
- Entry level to high end, unified user experience (EPU) – switch from tool to tool easily
- Robust connectivity through AutoLoader sample handling (AutoGrids)

- Outcome based support structure

- 165+ cryoEM field service and application specialists
- Time-to-result after install – apps support, EM schools

- Multiple ways to start with CryoEM

- Full suite- full control on your process and timing
- Through CryoEM service facilities- waiting list

Three-dimensional electron crystallography of protein microcrystals

Dan Shi[†], Brent L Nannenga[†], Matthew G Iadanza[†], Tamir Gonen*

Janelia Farm Research Campus, Howard Hughes Medical Institute, Ashburn, United States

Abstract We demonstrate that it is feasible to determine high-resolution protein structures by electron crystallography of three-dimensional crystals in an electron cryo-microscope (CryoEM). Lysozyme microcrystals were frozen on an electron microscopy grid, and electron diffraction data collected to 1.7 Å resolution. We developed a data collection protocol to collect a full-tilt series in electron diffraction to atomic resolution. A single tilt series contains up to 90 individual diffraction patterns collected from a single crystal with tilt angle increment of 0.1–1° and a total accumulated electron dose less than 10 electrons per angstrom squared. We indexed the data from three crystals and used them for structure determination of lysozyme by molecular replacement followed by crystallographic refinement to 2.9 Å resolution. This proof of principle paves the way for the implementation of a new technique, which we name **MicroED**, that may have wide applicability in structural biology.

DOI: [10.7554/eLife.01345.001](https://doi.org/10.7554/eLife.01345.001)

Single Particle Analysis (SPA)

- >100kDa particles
- Protein Complex
- Membrane Protein
- Best resolution 2-4Å

NEW

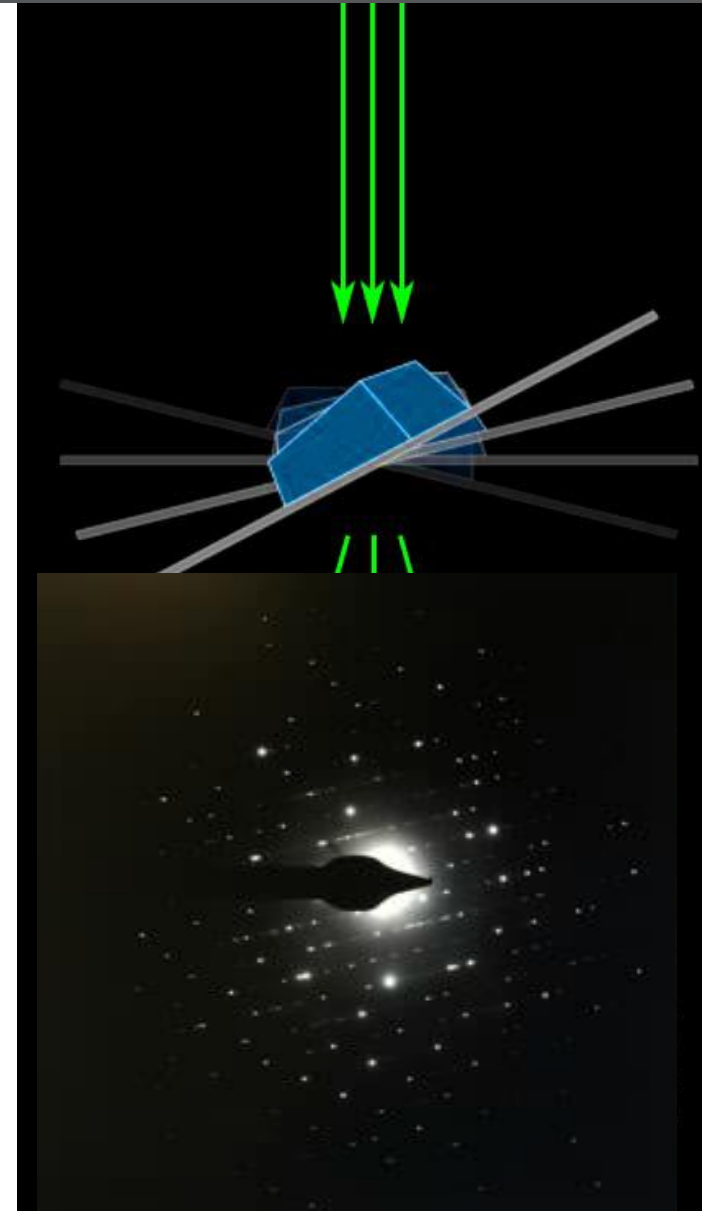
3D-Crystallography (MED)

- 1-200kDa particles
- Single proteins
- Pharma Molecules
- Best resolution 1-2Å

Introduction Micro Electron Diffraction

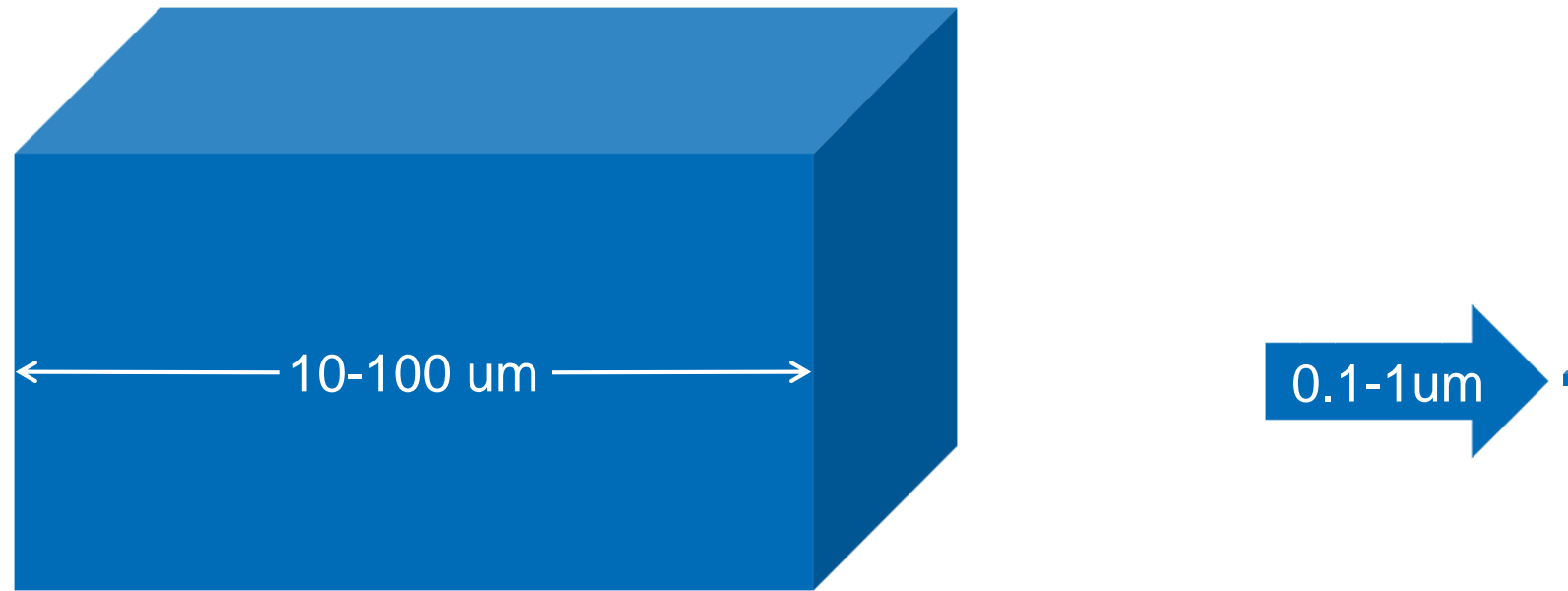
3D electron diffraction in a TEM on nano-crystals of bio-molecules using a continuous stage tilt

The 3D diffraction pattern can be used for high-resolution structure determination



Micro ED and X-ray crystallography

Micro Electron Diffraction can be done on crystal volumes which are a million times smaller than crystals used for X-ray crystallography.



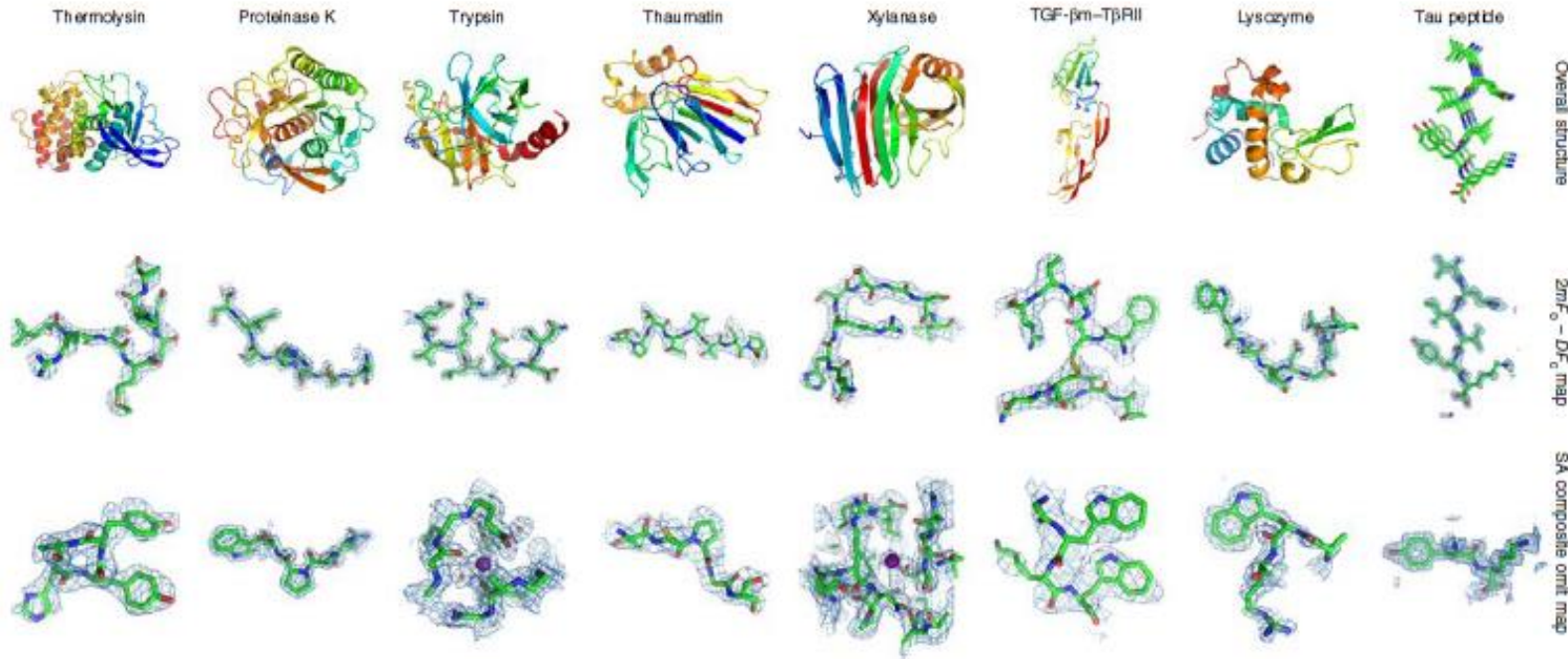
XFEL needs a bucket of nanocrystal to be injected in the system

MED needs between 2-10 nanocrystal

With a single image from a single crystal you can check if your crystal diffract nicely

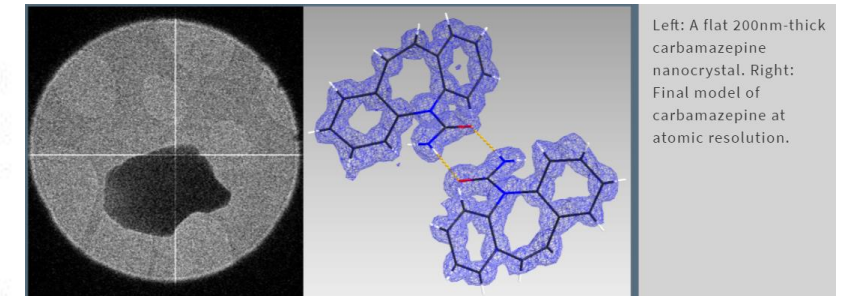
Main applications

Protein structure 1-3 Å resolution range

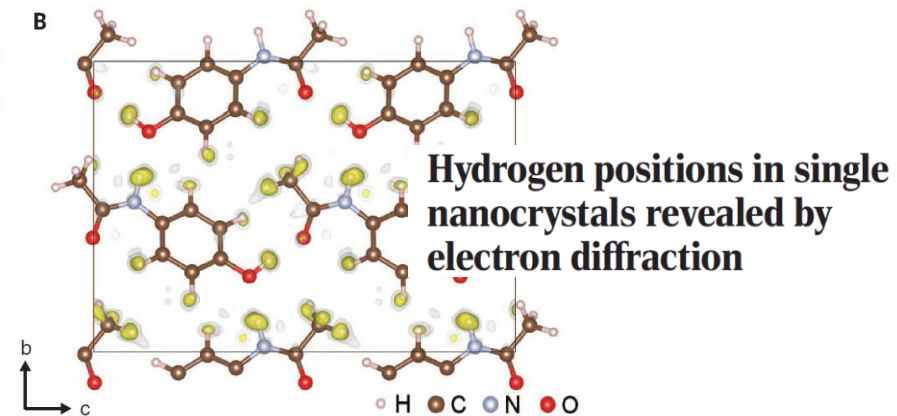


M. Jason de la Cruz *et al*, Atomic-resolution structures from fragmented protein crystals with the cryoEM method MicroED, Nature Methods 14, p.399 (2017)

Pharma molecules <1 Å resolution range

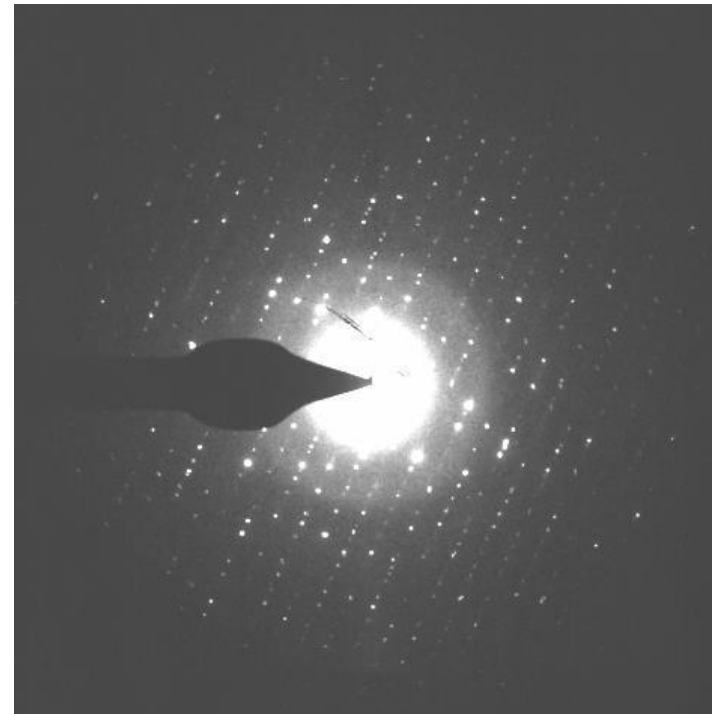


J.Abrahams *et al*, Carbamazepine
<https://c-cina.unibas.ch/research/ediff/roted/>



Palatinus *et al.*, *Science* 355, 166-169 (2017) 13 January 2017

- Thermo Fisher scientific is collaborating with Tamir Gonen (UCLA) on Micro Electron Diffraction to further pioneer this technique
- Testing work will be conducted on Talos Arctica
- System has an accurate single-tilt stage and is equipped with a modified Ceta camera (CMOS, scintillator based detector)



Join the resolution revolution!

Max Maletta
Max.Maletta@thermofisher.com

Cryo Tomography: Bridge protein structure with cellular function

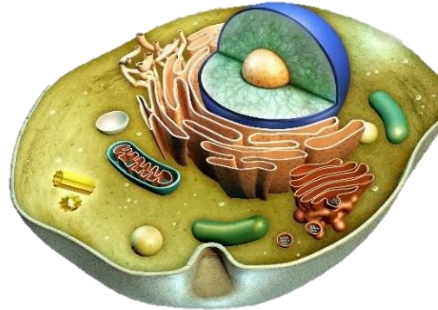
Protein coding Gene



Structure of
folded Protein



*Function of
Protein in cells*



*Cellular organisation
in tissue*

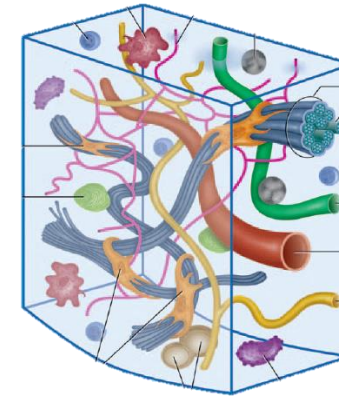


Illustration: Charis Tsevis (flickr)

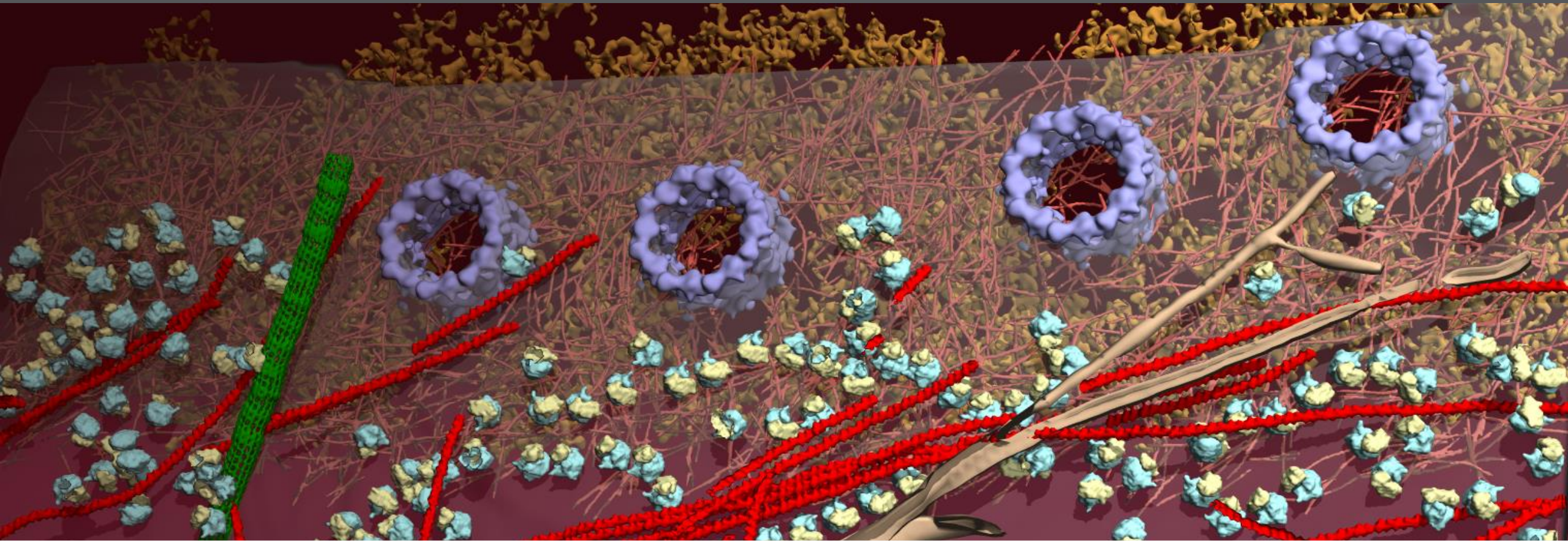
NextGen Sequencing

SPA

Cryo Tomography

Large Volume Analysis

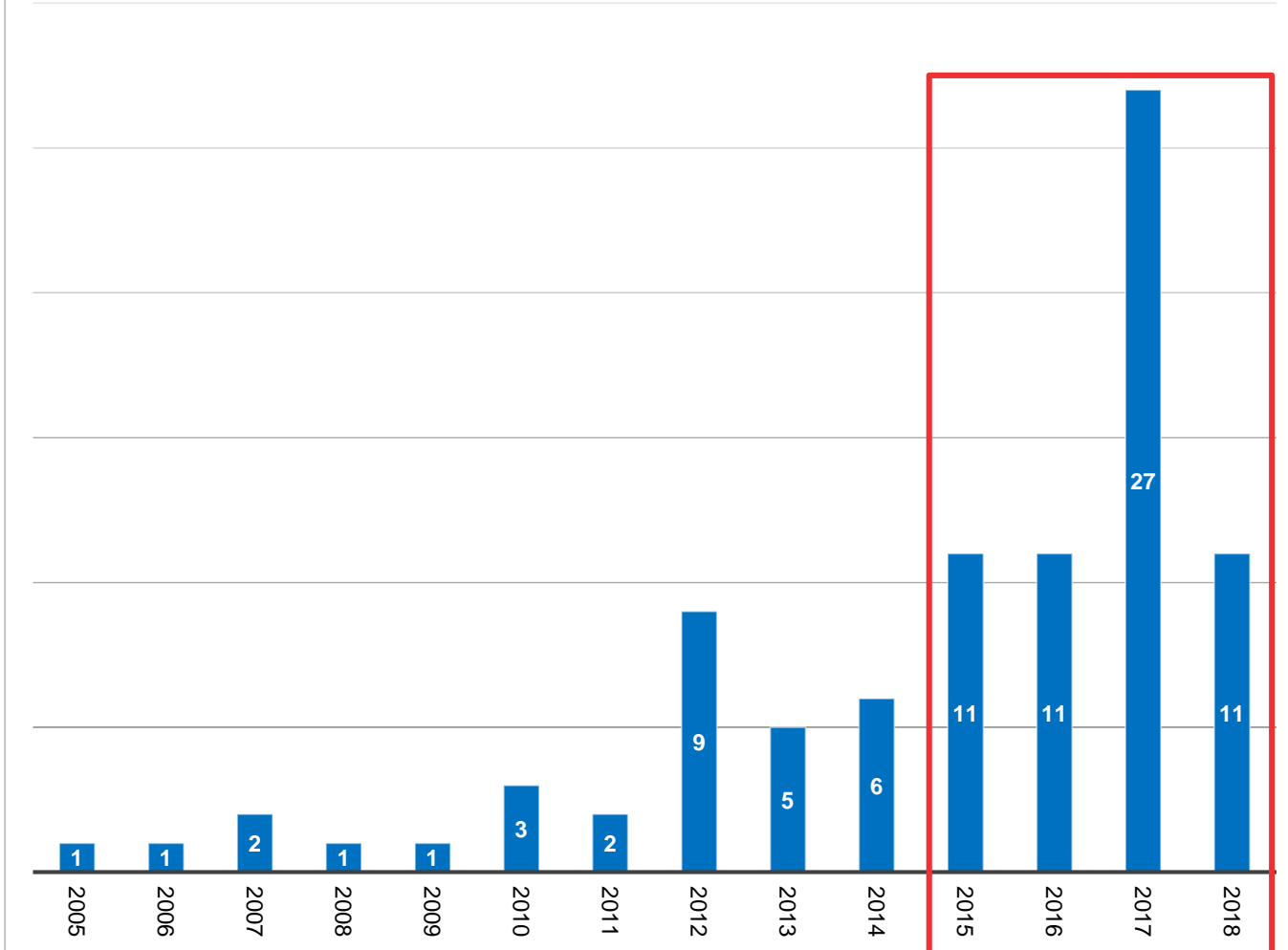
Cryo Tomography ...



- ... is the only technique that is non-disruptive to cellular components
- ... visualizes cellular structures at unprecedented resolution in 3D
- ... resolves structures of proteins inside the cell via sub-tomogram averaging
- ... allows us to understand entire processes inside cells

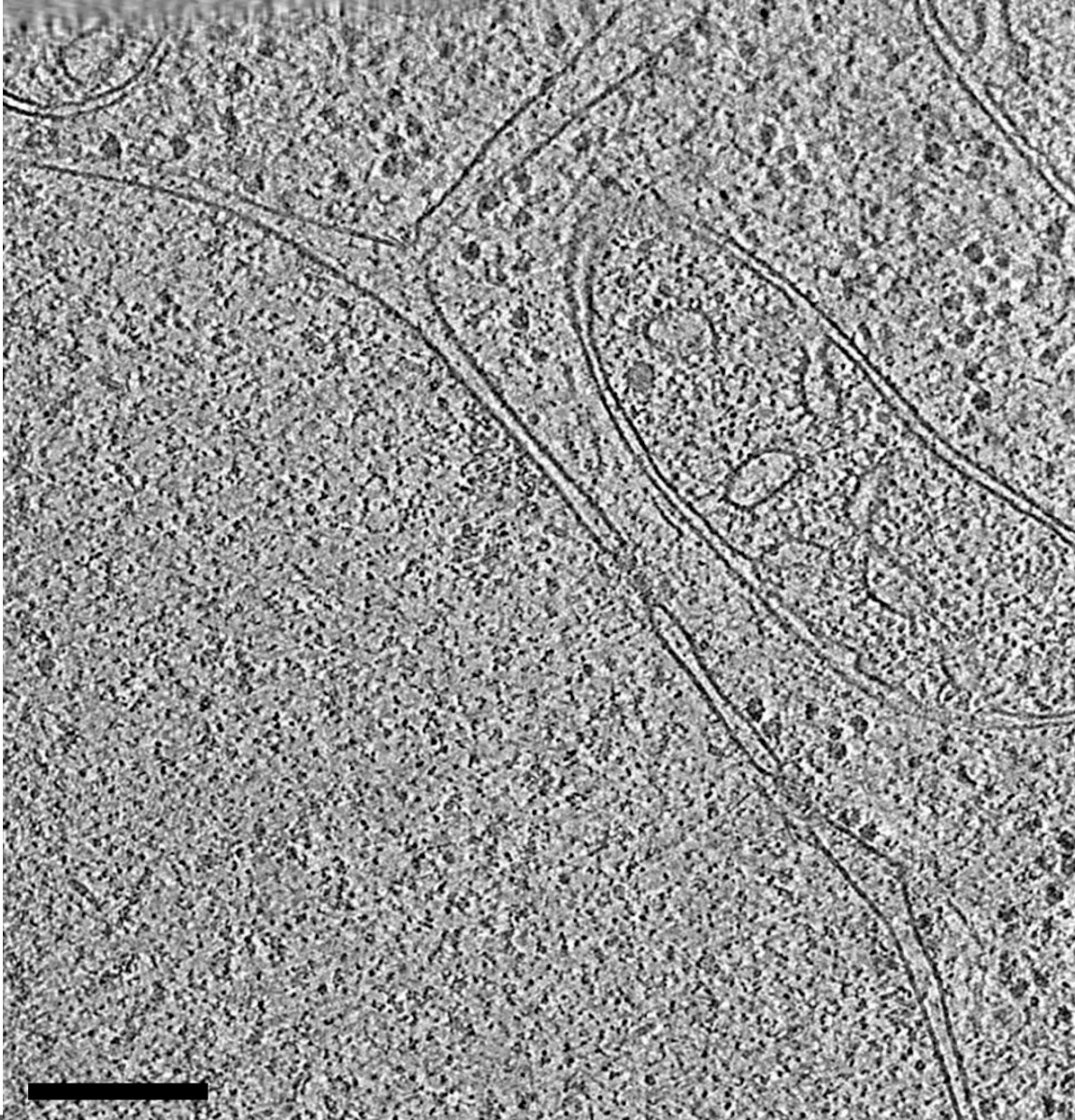
of publications are rising exponentially

Cryo Tomography Publications



- 60 Publications 2015-2018:
- 23% (14) in widely read journals like Cell, Science, Nature, PNAS.
- 86% Thermo Fisher Scientific Instruments

Proteasomes cluster around the NPC



The nuclear pore complex (NPC) selectively gates the transport of macromolecules between the nucleus and cytoplasm compartments

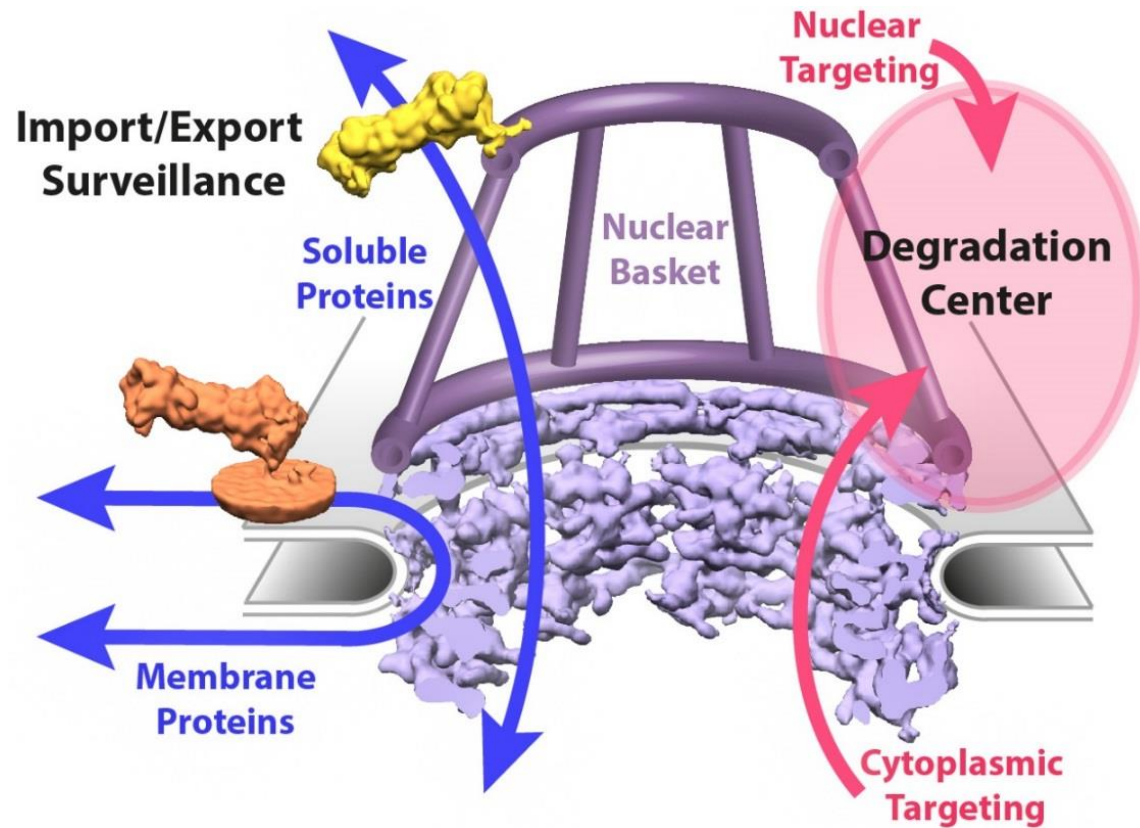
Research objective

Understand if surveillance mechanisms exist to reinforce the selective gate function

Unique

- *in situ* cryo-electron tomography enables to image the native cellular environment
- to perform an extensive molecular-resolution structural survey of cytoplasmic and nuclear proteasomes

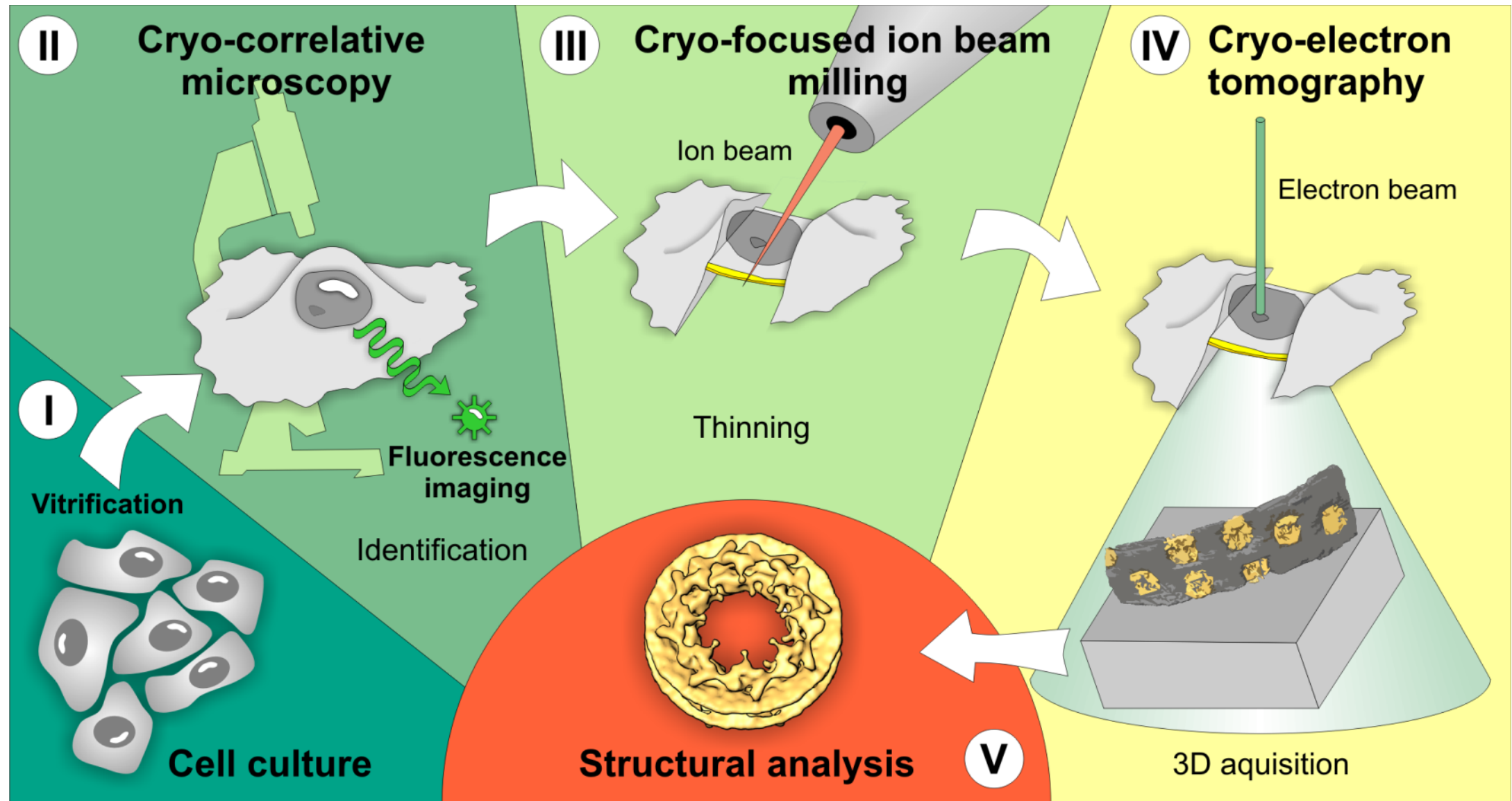
Proteasomes cluster around the NPC



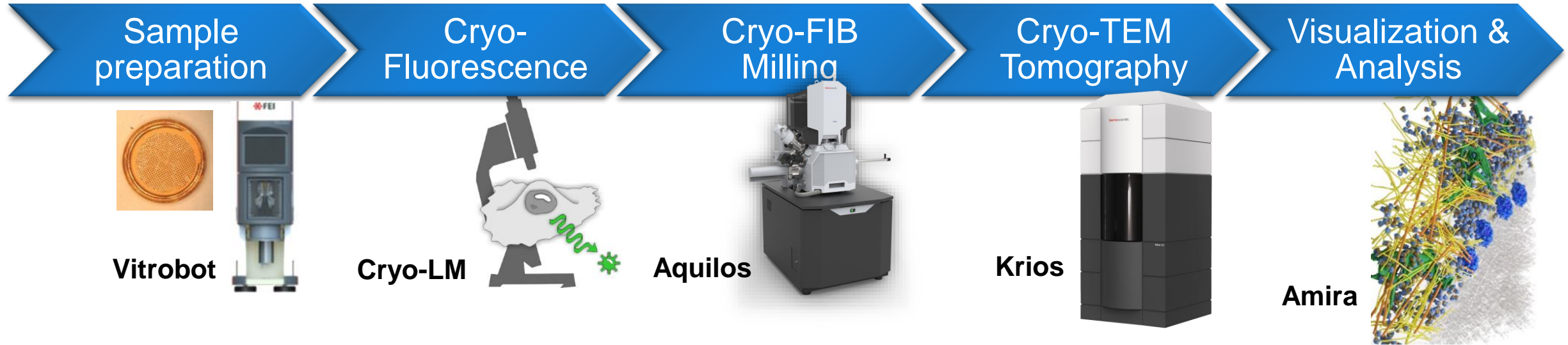
Results:

- the assembly states and functional states of proteasomes in each compartment were similar
- structural analysis revealed mechanistic details of the two NPC tethering interactions (nuclear basket and nuclear membrane)
- proteasomes bind to NPC establishing a cellular hub for protein degradation at the gateway between the nucleus and cytoplasm

Cryo Tomography Workflow: Enabling In Situ Structural Biology



Cryo-Tomography: Workflow challenges



Find area of
interest

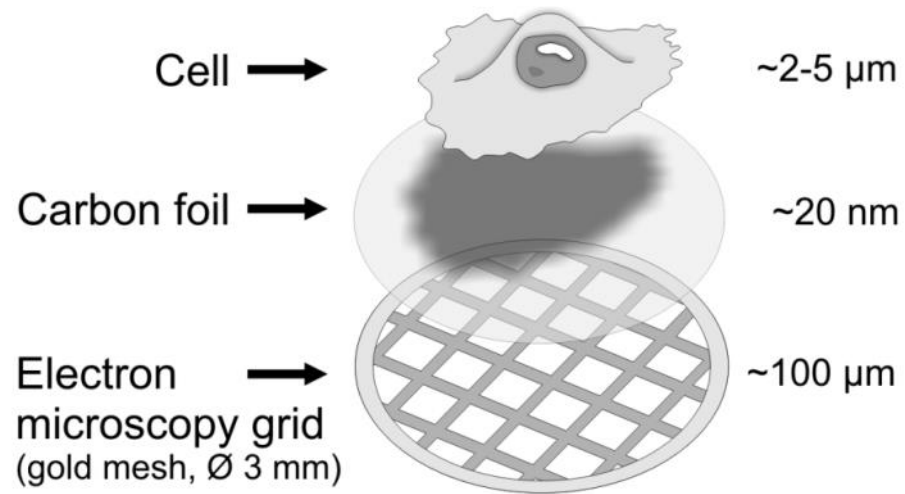
Easy connectivity
between
instruments

Lamella preparation fast
and with minimum
contamination

How to overcome the challenges?

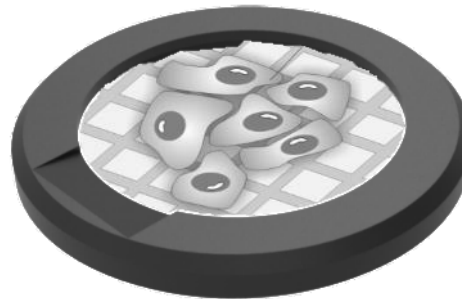
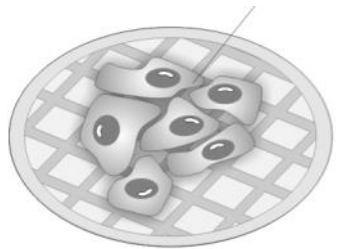
Sample Preparation and connectivity

Easy connectivity
between instruments



Dedicated Cryo-FIB autogrid

Vitrified cells on grid

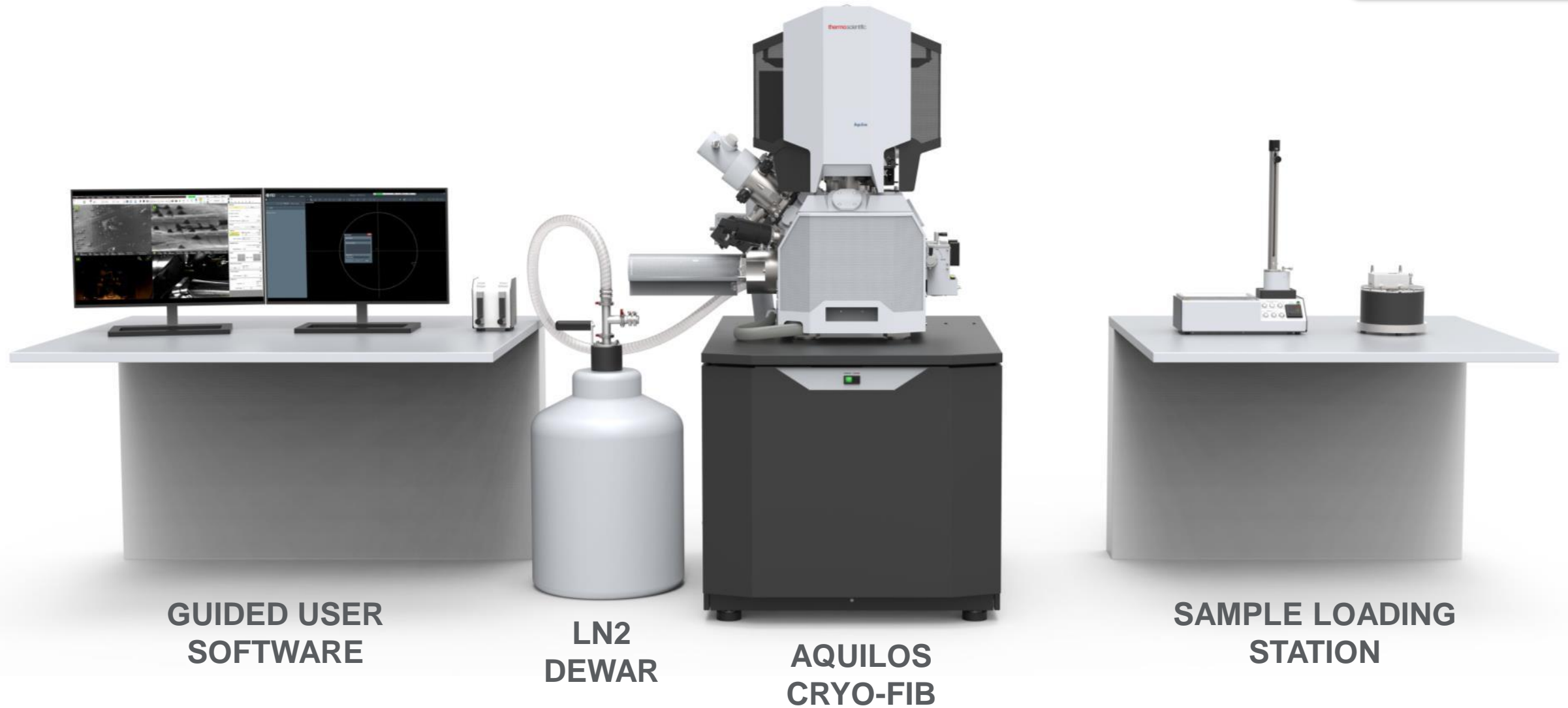


EM grid carrier/support

- Provides stability to fragile grids
- Allows robotic handling (FEI Autoloader) and automated TEM loading

Aquilos Cryo-FIB: A Complete Cryo-Sample Preparation Platform

Lamella preparation



Unique Value Propositions-Summary

- Fast adoption of Cryo-Tomo

- 30 publication in 2017 (total 60 from 2015)
- 23% (14) in high impact papers and 83 % based on Thermo Fisher instruments

- Enabling *in Situ* Structural Biology with fully dedicated platform

- Dedicated sample prep tool for cryo-tomography workflow
- Best compatibility and connectivity within Thermo Scientific's cryo tomography workflow
- Minimized artifacts
- Full control over thickness

- Workflow connectivity

- Robust connectivity to TEM through AutoLoader sample handling (AutoGrids)
- MAPS easy correlation and targeting software



Diamond Synchrotron Statistics

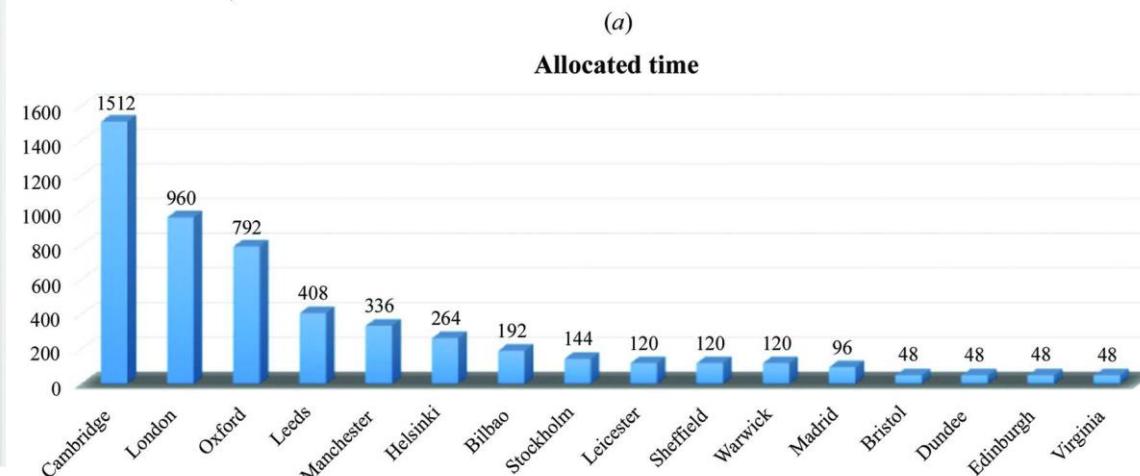
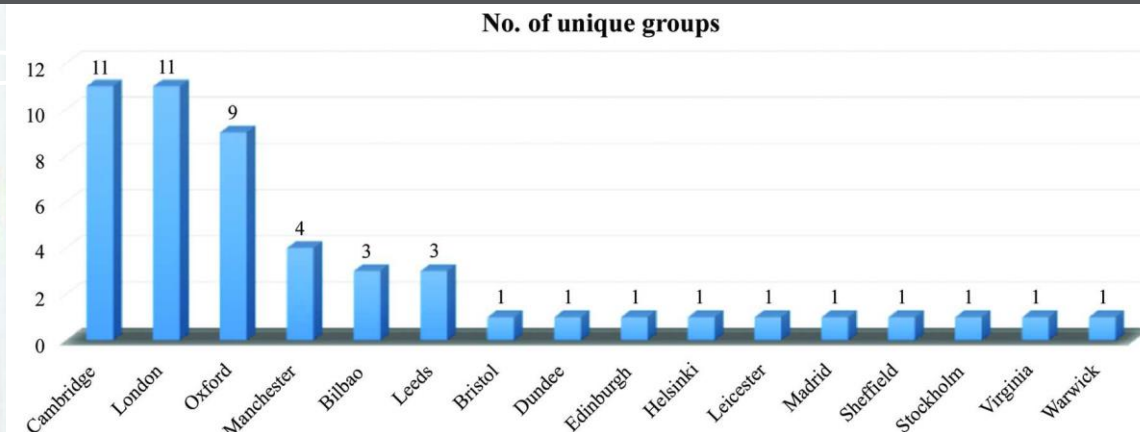
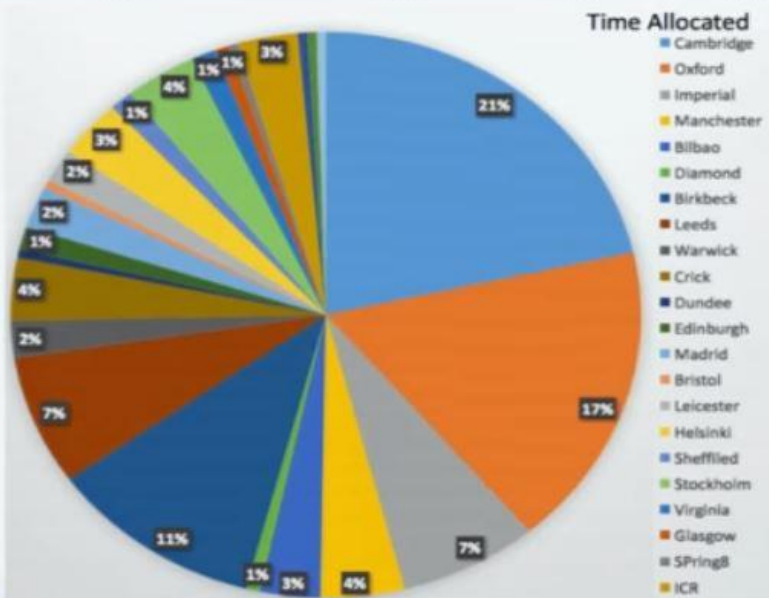
User statistics

Krios I&II use: 597 days (7/15 – 4/17, 1792 shifts)

| | Peer Reviewed | In House | Commissioning | Industrial | Training |
|-----------------|---------------|----------|---------------|------------|----------|
| Totals (shifts) | 1307.375 | 281.875 | 140.75 | 45 | 17 |
| Percentages | 73.0 | 15.7 | 7.9 | 2.5 | 0.9 |

As of 3/2017:

- 149 proposals awarded
- 216 sessions allocated (incl. BAGs)
- UK Users: 86%
- EU Users: 10.6%
- WW Users: 3.4%



(a)

- University of Oxford, and funded by the Wellcome Trust, the UK Medical Research Council (MRC) and the Biotechnology and Biological Sciences Research Council (BBSRC)

- Typical imaging time 48hrs
- 95 Separate Visits / 53 Individual Investigators
- 18 Total publications as of 2/2018

- First year of operation generation of 270TB of data via Krios 1
- Estimation will be up to 1 PB of data as Krios 4 comes online