Enzymatic degradation of poly(3-hydroxybutyrate): qualitative analysis and kinetical representation

///GE

Péter Polyák 1,2, Emese Dohovits1, Gergely Nagy3,4, Beáta Vértessy3,4, Béla Pukánszky1,2

Laboratory of Plastics and Rubber Technology, Department of Physical Chemistry and Materials Science, Budapest University of Technology and Economics ²Institute of Materials and Environmental Chemistry, Research Centre for Natural Sciences

³Department of Applied Biotechnology and Food Science, Budapest University of Technology and Economics

⁴Laboratory of Genome Metabolism and Repair, Institute of Enzymology, Research Centre for Natural Sciences

Background

In order to make the future of polymer industry sustainable, we have to eliminate the need for fossil resources, as these resources are inevitably depleting, on the one hand, and the carbon footprint of polymers produced from fossil material sources is enormous, on the other. A possible solution to these drawbacks is offered by the results of research projects conducted in the field of biopolymers: these materials can now be used as potential substituents of conventional polymers, e.g. polyolefins. Moreover, these materials can also biodegrade, which makes their recycling fast, easy and cost efficient. These advantageous features are originating from the fact that the depolymerization of the vast majority of these polymers can be catalyzed by enzyme molecules.

Aim of the work

The goal of the research is to investigate the degradation of poly(3-hydroxybutyrate) (PHB) initiated and catalyzed by enzyme molecules natively produced by the strain Bacillus Megaterium, and also to describe the reaction with a modified Michaelis-Menten model quantitatively. Enzyme molecules are expressed in the Department of Applied Biotechnology and Food while the experiments aiming to characterization of depolymerization are executed in the Laboratory of Plastics and Rubber Technology.

Methods

A) Enzyme expression with a recombinant Escherichia coli strain

Recombinant bacteria:

Strain:

Origami DE3 (Novagen) pGS1865 (pET22b+, bearing the Bacillus megaterium intracellular PHB depolymerase gene)

Enzyme separation and purification:

Method Column affinity chromatography Ni-NTA Agarose

B) Amorphous PHB film preparation

Temperature

Compression molding

120 kN Pressure

C) Enzymatic degradation monitored by UV-VIS spectrophotometry

Instrument Unicam UV-500

PharmaTest PTWS 600

Resolution

190-400 nm Wavelength range

D) Enzymatic degradation monitored by liquid chromatography

Chromatograph Eluent:

Wavelength range

Merck-Hitachi LaChrom Elite pH 3.0 H₃PO₄/KH₂PO₄ buffer 1 ml/min

Flow:

LiChroChart 250-4 LiChrospher 100 RP-18 5 μm DAD UV-VIS Stationary phase: Detector

190-400 nm

Materials





Degradation product: 3-hydroxybutyric acid

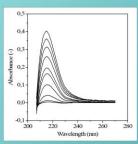
Results

Secondary and tertiary structure of a PHB depolymerase enzyme

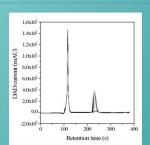
1) Increasing absorbance at 215 nm

Both the UV-VIS and HPLC-DAD measurements measurements show an increasing absorbance at 215 nm, which can be increasing attributed to the formation of metabolite molecules.

The monomer, (3-hydroxybutyric acid) proved to be the sole degradation product by the recursively recorded HPLC chromatographs.



Time dependent UV spectra of the aqueous media

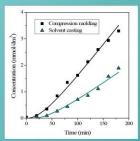


Time dependent chromatograms of

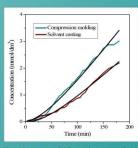
2) Time dependent monomer concentration

function has is nonlinear. The enzymatic reaction shows a slightly accelerating characteristic at the beginning, which is later followed by a stationary phase

The explanation of the nonlinear reaction characteristics shown by the graphs presented here requires the application of a kinetic model



Absorbance values at the UV-VIS spectrum peak maximums



Absorbance at 215 nm (measured with an online UV-VIS spectrophotometer)

3) Kinetic characterization based on the modified Michaelis-Menten model

modification consideration the heterogeneous nature of the enzymatic PHB degradation

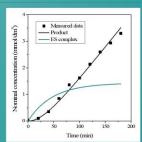
$$\frac{d[ES](t)}{dt} = +k_1 \cdot E_0 \cdot S_0 - k_{-1} \cdot [ES](t)$$
$$-k_{cat} \cdot [ES](t)$$

$$\frac{d[P](t)}{dt} = +k_{cat} \cdot [ES](t)$$

An analytical solution of the provides the [P](t) function looked

$$P(t) = C \cdot \frac{-k_{cat}}{(k_{-1} + k_{cat})} \cdot e^{-(k_{-1} + k_{cat})}$$
$$+ k_{cat} \cdot \frac{k_1 \cdot E_0 \cdot S_0}{k_{-1} + k_{cat}} \cdot t + C'$$

An adequate fitting of the model onto the measured data provides the reaction rate coefficients which can be applied to predict time dependence product concentration.



Conclusions

- 1. The intracellular PHB depolymerase of the strain Bacillus megaterium is an excellent catalyst of enzymatic degradation.
- 2. HPLC measurements prove that the sole product of the reaction is the monomer (3-hydroxybutyric acid).
- 3. Monomer formation can be described with, or even predicted by a modified Michaelis-Menten model developed in the project.